

CHANGES IN PRODUCTIVE PERFORMANCE, HEMATO-BIOCHEMICAL INDICES, IMMUNE AND ANTIOXIDANT STATUS OF GROWING OSSIMI LAMBS SUBJECTED TO VITAMINS A AND E ADMINISTRATION

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ABSTRACT

This study included 32 growing Ossimi male lambs, averaged 3.5 months of age and 19.08 ± 1.09 kg body weight. They divided into four equal groups (8 lambs each) to determine the effects of vitamins A and E oral administration on growth performance, hematological and biochemical parameters, immune and antioxidant status. The 1st group served as control, the 2nd group (VA) received oral administration of vitamin A at 50,000 IU/head/biweekly, the 3rd group (VE) received vitamin E at 400 mg/head/biweekly, and the 4th group (VA+E) received 50,000 IU vitamin A plus 400 mg vitamin E /head/biweekly. Data showed an increase ($P < 0.05$) in body weight (BW) of lambs received VA and VA+E vs. control. Average daily gain (ADG) increased ($P < 0.05$) for lambs received VA and VA+E vs. control and VE treatment. ADG was higher ($P < 0.05$) for lambs received VA+E than those received each of VA or VE alone. Feed conversion efficiency (FCE) was improved ($P < 0.05$) for lambs received VA and VA+E vs. control and VE treatment. No differences in dry matter intake (DMI) was recorded among the experimental groups. Blood Hb concentrations increased ($P < 0.05$) for lambs received VA, VE and VA+E, while RBCs count and PCV % were increased ($P < 0.05$) only for lambs received VE and VA+E. No significant response of WBCs count among lambs received VA, VE and VA+E vs. control. Lymphocytes % increased ($P < 0.05$) with no significant differences in eosinophils, basophils and monocytes % for lambs received VA, VE and VA+E vs. control. Lambs treated with VE alone exhibited a decrease ($P < 0.05$) in neutrophils vs. lambs of control, VA and VA+E. Serum total protein (TP) and globulin concentrations increased ($P < 0.05$) with VA, VE and VA+E treatments vs. control. Serum TP and albumin concentrations increased ($P < 0.05$) for lambs treated with VA+E treatment when compared to those treated with VA treatment alone. Lambs received VA, VE and VA+E treatments had higher ($P < 0.05$) serum IgG concentrations than the control. VE treatment increased ($P < 0.05$) serum IgG concentrations vs. VA treatment. There were significant ($P < 0.05$) increases in serum total antioxidant capacity and glutathione peroxidase (GSH-Px) activity for lambs received VA, VE and VA+E treatments vs. control. Lambs of VE treatment had higher ($P < 0.05$) serum GSH-Px activity than those of VA treatment. No significant differences in serum concentrations of glucose, cholesterol, AST and superoxide dismutase activity was noticed due to treatments VA, VE and VA+E vs. control. These results indicate that combination of vitamins A and E exerted beneficial additive effects that improve ADG and physiological responses of lambs. Vitamin A was more effective than vitamin E in enhancing growth performance of lambs, whereas vitamin E had more potent effect on improving immune response and antioxidant status.

Key words: Vitamin A and E, Growing Lambs, Performance, Physiological responses, Antioxidant status.

INTRODUCTION

Antioxidant vitamins are vital components for mammalian cell defense against compounds that cause oxidation of cellular molecules. Vitamins A and E are essential, natural occurring and fat-soluble nutrients that are involved in several important biological processes such as immunity, protection against

tissue damage, reproduction, growth and development (**Debier and Larondelle, 2005**). Vitamin A is essential for stimulation of growth, proper development of skeletal tissue, normal reproduction and maintaining the integrity of epithelial tissues (**Pond et al., 1995**). Vitamin E, the most potent antioxidant, works to scavenge free radicals and acts as

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terminator of lipid peroxidation (Liebler, 1993). Vitamins A and E are cellular antioxidants, preventing peroxidative damage in cell membranes, and are essential for the well-functioning immune system (Meglia *et al.*, 2004). Animal immune function and health could be impaired by inadequacies of vitamin A and E as antioxidant defense (Chew, 1987). Therefore, the amount of vitamin A needed for immune-enhancement is reported to be higher than the suggested required amounts by NRC (Nockles and Blair, 1996). Supplement with vitamin E at levels above requirements is also associated with variable improvements in sheep performance and immune function (Rooke *et al.*, 2004). Vitamin E is considered synergistic to vitamin A due to its antioxidant activity, which results in a sparing effect upon vitamin A. Therefore, loss of vitamin A from liver stores is accelerated in the presence of vitamin E deficiency, resulted of increased fragility of lysosomal membranes (Watts, 1991). Vitamin E therapy appears to be an effective treatment for hypervitaminosis of vitamin A (St-Claire *et al.*, 2004). In cattle, high dietary levels of vitamin A could depress vitamin E utilization (Schelling *et al.*, 1995). In sheep, however, it was suggested that oversupply of dietary vitamin A does not antagonize vitamin E turnover (Hidiroglou, 1993); and that daily administration of vitamin A, approximately 150 times greater than the daily requirement, were well tolerated by sheep (Raofi *et al.*, 2010). In rats, it has been suggested that vitamin A might be considered as a potential antioxidant similar to vitamin E in animal nutrition (Kantha and Krishnamurthy, 1977).

However, the inter-relationships between vitamins A and E are complex and the mechanisms of their interaction are not fully understood. The present study, therefore, focused on some mechanistic aspects through which administration of vitamin A and vitamin E may influence growth performance of growing Ossimi lambs and their physiological reactions related to hematology, serum biochemical profile, immune and antioxidant status.

MATERIAL AND METHODS

Animals and experimental treatments:

Thirty-two of growing Ossimi male lambs average 3.5 months age and 19.08±1.09 kg body weight were used in this study during November 2013 to February 2014 at the Farm of Animal Production Department, Faculty of Agriculture, El-Minia University. The animals were randomly divided into 4 equal groups (8 lambs each) of similar initial body weights. The 1st group served as control, the 2nd group (VA) received oral administration (drenching) of vitamin A (as palmitate) at rate 50,000 IU/head/biweekly, the 3rd group (VE) received vitamin E at rate 400 mg/head/biweekly, and the 4th group (VA+E) received vitamin A at 50,000 IU/head/biweekly plus vitamin E at 400 mg/head/biweekly.

Feeding and management:

Animals were fed on concentrate feed mixture (CFM) and rice straw to cover their nutrient requirements according to live body weight (NRC, 1985). The CFM consisted of 15 % yellow corn, 15 % soybean meal, 30 % sugar beet pulp, 37 % wheat bran, 2.0 % limestone and 1.0 % salt. The calculated feeding value of the CFM was 69.75 % TDN, 17.04 crude protein and 2.53 ME (Mcal/kg). The concentrate contained 1.41 mg/kg DM β -carotene and 11.97 mg/kg DM vitamin E. The NRC requirements for growing lambs are 69 μ g of β -carotene/kg live weight/day (47 IU of vitamin A/kg live weight/day) and 20-25 mg of vitamin E /lamb/day. The treated animals received 1.41 mg/kg DM of dietary β -carotene with 50,000 IU/head/biweekly of vitamin A and 11.97 mg/kg DM with 400 mg/head/biweekly of vitamin E. Feeds were offered twice a day at 8 am and 2 pm and mineral blocks and drinking water were available along the experiment. Body weights of lambs were recorded at the start of experiment then biweekly. Feed intake was recorded daily. Averages of daily gain and feed conversion efficiency of lambs were calculated. Parameters were recorded in the morning before animals access to feed or water.

Blood sampling and measurements:

Heparinized blood samples were collected from the jugular vein of each animal at 8.00 am before feeding or drinking. Whole blood samples were analyzed after collection for hemoglobin (Hb), packed cell volume (PCV), red blood cell counts (RBCs) and white blood cell counts (WBCs). The Hb concentration was determined using cyanomethomoglobin method (Campbell, 1995). The PCV was determined using micro-hematocrit tubes with micro-hematocrit centrifuge at 12000 rpm for three minutes. The RBCs and WBCs were counted using the light microscope. Stained blood smears with Lieshman's stain were prepared for the differential WBCs count (Dacie and Lewis, 1991). Non-heparinized blood samples were collected from the jugular vein of each animal and left to clot at room temperature for at least 4 h, then the clots were removed and sera were cleared by centrifugation at 1500×g for 20 min and stored at -20 °C for later biochemical assay. Serum glucose, total protein, albumin, cholesterol and aspartate transaminase (AST) were determined colorimetrically using Bio-diagnostic product kits (Egypt). Serum globulin concentrations were calculated by difference between total protein and albumin concentrations. Serum immunoglobulin G (IgG) was quantified using ELISA kit supplied by WKEA MED Supplies Corporation. The Elisa micro plate having standards (5 wells) and samples was read at 450 nm using Elisa READER (BIO TEK ELX808), USA. The assay of serum IgG range was 0.7 to 30 µg/ml. Serum total antioxidant capacity (TAC), superoxide dismutase (SOD) and glutathione peroxidase (GSH-Px) activities were analyzed colorimetrically by STAT LAB SZSL60-SPECTRUM, using Bio-dignostic kits (Bio-dignostic Company, Egypt). The analyses were performed at Cairo University Research Park (CURP), Faculty of Agriculture, Cairo University.

The data were analyzed by least square means analysis of variance using General Linear Models (GLM) procedure of the statistical analysis system (SAS, 2000). The model used to analyze different traits studied for ewes or lambs was as follows:

$$Y_{ij} = \mu + T_i + e_{ij}$$

Where: Y_{ij} = i th Observation, μ = Population mean; T_i = Effect of i th treatments and e_{ij} = Random error. Duncan's Multiple Range test was used to detect differences between means of the experimental groups (Duncan, 1955).

RESULTD AND DISCUSSION

Productive performance:

The results in Table (1) illustrate the effect of vitamin A (VA) and/or vitamin E (VE) on growth performance of Ossimi growing lambs. Data showed a significant ($P < 0.05$) increase in final body weight (FBW) of lambs received VA and VA+E vs. control and the higher FBW was recorded with VA+E treatment (38.76 kg). Average daily gain (ADG) increased ($P < 0.05$) by 40.3 and 69.8 % respectively for lambs received VA and VA+E vs. control. ADG was higher ($P < 0.05$) for lambs received VA+E (219.0 g/d) than those received each of VA (181.0 g/d) or VE (145.0 g/d) alone. Feed conversion efficiency (FCE) was improved ($P < 0.05$) by 38.2 and 69.0 % for lambs received VA and VA+E vs. control. No significant differences in dry matter intake (DMI) were recorded among the experimental treatments.

The significant improvement in FBW, ADG and FCE for lambs received VA are in line with some earlier studies showed significant beneficial effects of vitamin A that improve productive performance of growing calves (El-Masry *et al.*, 1998), sheep (Bruns and Webb, 1990; Soliman, 2005) and buffalo calves (El-Barody *et al.*, 1993). Since vitamin A has major function in metabolism that preserve stability, structural integrity and normal permeability of cell and subcellular membranes, it has positive effects on tissue biosynthesis and growth promotion (Chew, 1993). Generally, vitamin A is supplemented to ruminant especially to those confined to insure their optimum health and maximum productivity (Alosilla *et al.*, 2007). Vitamin A has a role in regulating growth hormone gene expression (Bedo *et al.*, 1989), and energy homeostasis by enhancing uncoupling protein 1 (UCP1) mRNA gene expression and decreasing

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serum leptin levels (Kumar *et al.*, 1999). Thus, metabolic disorder, reduced feed efficiency, slowed gains and growth retardation are clinical signs occurred with lack of vitamin A (Pond *et al.*, 1995). Although the effect of vitamin E on lambs' performance was statistically not significant, the averages of FBW, ADG and FCE tended to improve by 4.8, 12.5 and 8.5 % for lambs received VE vs. control (Table, 1). Though supplement of vitamin E was beneficial in improving animal' performance of lambs (Shetaewi *et al.*, 1992), buffalo calves (Amer and Hashem, 2008) and cattle calves (Hays *et al.*, 1987 and Galyean *et al.*, 1999), other reports showed no significant or limited responses in performance' parameters with supplement of vitamin E on growing male lambs (Zhao *et al.*, 2013), goats (Yang *et al.*, 2010) and beef cattle (Rivera *et al.*, 2002).

The presented results clearly show that co-administration of VA+E in lambs were more effective and significantly increased their ADG than each of VA or VE alone (Table, 1). In addition, the combined treatment (VA+E) also improved FCE vs. control or VE-treatment alone. These results may signify the additive effect of vitamin A and E that enhance lambs' performance. Such positive combined effect of both vitamins was shown in Holstein steer calves received 30,000 IU/d of vitamin A plus 250 IU/d of vitamin E which improved (P<0.05) their FBW and ADG and slightly enhance DMI with no significant effect on gain or dietary net energy (Salinas-Chavira *et al.*, 2014). Furthermore, a combination of vitamins

A and E was more effective than either vitamin alone in reducing heat stress performance in broiler (Sahin *et al.*, 2001). The relationship between vitamin A and vitamin E has been proposed in such a way that vitamin E appears to have an important effect on the utilization and perhaps absorption of vitamin A and that vitamin E protects vitamin A from oxidative breakdown (Gallo-Tores, 1980). In contrast, dietary supplementation of vitamin A had no effect on performance (Feed intake, ADG and FBW) of young lambs (Arnett *et al.*, 2007). Several factors including the extent and duration of treatment, age, phase of growth, species, route of administration and other environmental conditions, likely contribute to the discrepancies in growth performance response to both vitamins supplementation.

Hematological parameters:

The results in Table 2 showed that hematological parameters (Hb, RBCs and PCV) were significantly changed in response to vitamin A and/or vitamin E administration. Blood Hb concentration increased (P<0.05) for lambs received VA, VE and VA+E, while RBCs and PCV % were increased (P<0.05) for lambs received VE and VA+E than control or VA treatment. This response of hematologic indices may signify a case of active metabolism and biological oxidation on the cellular base for these treated lambs (Frandsen, 1986) that lead to availability of metabolites required for tissue growth.

Table 1: Effects of vitamin A and vitamin E administration on productive performance of growing Ossimi lambs (mean ± SEM).

Parameters	Treatments				SEM	Sig.
	Control	VA	VE	VA+E		
IBW (kg)	19.08	19.08	19.08	19.08	1.09	NS
FBW (kg)	30.65 c	35.38 ab	32.11 bc	38.76 a	1.47	*
ADG (g/day)	129.0 c	181.0 b	145.0 c	219.0 a	0.008	*
DMI (kg/head/day)	1.14	1.20	1.18	1.22	0.045	NS
FCE (kg feed/kg gain)	9.2 a	6.66 b	8.49 a	5.54 b	0.59	*

a,b,c means within the same row having different superscripts significantly different

(* P<0.05), NS = not significant

VA = Vitamin A, VE = Vitamin E, VA+E = Vitamin A plus vitamin E.

IBM = Initial body weight, FBW = Final body weight, ADG= Average daily gain,

DMI = Dry matter intake, FCE = Feed conversion efficiency.

Table 2: Effects of vitamin A and vitamin E administration on hematology and differential leucocytes count parameters of growing Ossimi lambs (mean \pm SEM).

Parameters	Treatments				SEM	Sig.
	Control	VA	VE	VA+E		
Hb (g/dl)	10.04 c	11.58 a	12.72 b	12.15 ab	0.30	*
RBCs ($\times 10^6/\text{mm}^3$)	9.0 b	9.6 b	11.4 a	11.1 a	0.21	*
PCV (%)	29.0 b	30.1 b	35.7 a	33.8 a	1.04	*
WBCs ($\times 10^3/\text{mm}^3$)	7.22	7.24	7.30	7.44	0.20	NS
Neutrophils (%)	30.66 a	28.1 a	25.0 b	27.8 ab	2.40	*
Eosinophils (%)	4.06	4.18	4.04	4.1	0.15	NS
Basophils (%)	0.55	0.52	0.51	0.5	0.02	NS
Lymphocytes (%)	58.8 b	63.1 a	66.2 a	63.3 a	3.77	*
Monocytes (%)	4.00	4.10	4.15	4.0	0.11	NS

a,b,c means within the same row having different superscripts significantly different

(* $P < 0.05$). NS = not significant

VA = Vitamin A, VE = Vitamin E, VA+E = Vitamin A plus vitamin E.

Some studies reinforce the present results showing similar response of increasing blood Hb due to vitamin A administration in sheep (Soliman, 2005), goats (Yang *et al.*, 2010) and Holstein dairy calves (Moosavian *et al.*, 2010). Vitamin A appear to be involved in the pathogenesis of anemia through diverse biological mechanisms via enhancing growth and differentiation of erythrocyte progenitor cells, potentiates immune system to infection, reduces the anemia of infection and requiring it for Fe metabolism and mobilization (Semba and Bloem, 2002). Moreover, studies in rats have shown that Fe deficiency alters plasma and liver levels of vitamin A. So, vitamin A deficiency may associated with altered Fe metabolism, including reduced plasma Fe and sometimes anemia; and this effect does not appear to be caused by increased RBC destruction (Pond *et al.*, 1995). Indeed, evidences in dairy calves treated with vitamin A and/or Fe indicated that the relationship between vitamin A and Fe remains to be unclear (Moosavian *et al.*, 2010). The significant increase in Hb, RBCs and PCV for lambs treated with VE has similar response, for these parameters, to vitamin E supplement found in coarse-wool lambs (Shetaewi *et al.*, 1992). Hematological responses to vitamin E could mediate its enhancing for erythropoiesis and decreasing the premature erythrocyte hemolysis by reducing the fragility of erythrocytes (Jiliani and Iqbal, 2011).

As shown in table 2, no significant response of WBCs count was observed among lambs received VA, VE and VA+E compared to control. The WBCs profile showed a marked increase ($P < 0.05$) in lymphocytes (%) concomitant with no significant differences in eosinophils, basophils and monocytes percentages for lambs received VA, VE and VA+E vs. lambs of control, but they did not show significant differences among the treated groups. However, lambs treated with VE alone exhibited a decrease ($P < 0.05$) in neutrophils vs. lambs of control, VA and VA+E. The insignificant change in WBCs count in lambs treated with VA agrees with similar response found in sheep (Soliman, 2005), goats (Yang *et al.*, 2010) and dairy calves (Moosavian *et al.*, 2010). The response of significant increase in lymphocyte percentages for lambs treated with VA, VE and VA+E may be considered a useful response that improve their immune function, disease resistance and general health. The increase in blood lymphocyte populations may be a good indicator of an immune response (Qureshi *et al.*, 2001). The presented results are consistent with some studies dealt with the effect of vitamins A or E on lymphocyte counts. For instance, lymphocyte populations were significantly increased in response to either injection of vitamin E in calves (Reddy *et al.*, 1986) and in Awassi rams (Ali *et al.*, 2009), or dietary supplementation of vitamin A in goats (Yang *et al.*, 2010), indicating that both

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vitamins could improve the immune function of these animals. Vitamin E can affect lymphocyte production in the bone marrow of ruminants (Hogan *et al.*, 1993). Vitamin A also plays a central role in the development and differentiation of WBCs, such as lymphocytes, which play critical roles in immune response (Semba, 1998).

In the current study, VA and VA+E treatments had no significant effect on neutrophils %, while VE treatment significantly reduced neutrophils % with no significant differences in the % of monocytes, acidophils or basophils (Table, 2). Similar results were noticed in Awassi rams when neutrophils % were decreased with the injection of vitamin E (Ali *et al.*, 2009). However, neither phagocytic index nor neutrophils % was affected by vitamin E injection in cattle (Hogan *et al.*, 1992). In *in vitro* study on growing calves, Eicher *et al.* (1994) reported that neutrophil phagocytosis improved with vitamins A and E compared to vitamin E alone. Indeed, differences in neutrophils response affected by many factors including supplement levels of vitamin E, mode of administration, selenium status and animal species used. Monitoring the hematological indices in sheep gives a clear picture of their nutritional and health status before the changes are visible on the animal (Antunovic *et al.*, 2009).

Serum biochemical parameters:

Data in Table 3, showed an increase ($P < 0.05$) in serum total protein (TP) and globulin concentrations with VA, VE and VA+E treatments vs. control while serum TP and albumin concentrations increased ($P < 0.05$) for lambs treated with VA+E compared to those treated with VA alone. This result may revealed that administration of vitamin A and/or vitamin E at dosage used in this study improved protein synthesis and metabolism. This response may be account for the trend towards improved growth performance (FBW, ADG and FCE) for these treated lambs. The present results agree with comparable responses reported by El-Shahat and Abdel-Monem (2011) who demonstrated that dietary supplementation of vitamin E significantly improved levels of

serum TP, albumin and globulin concentrations in Baladi sheep. Elevated plasma globulin was observed on lambs received VA as shown in Table 3. El-Masry *et al.* (1998) noticed similar trend of increasing globulin fractions for growing calves supplemented with vitamin A, but they failed to show a significant increase in plasma TP or albumin. In Ossimi sheep, supplement with vitamin A significantly increased plasma TP and albumin, however, globulin was not affected (Soliman, 2005). In cattle, feeding vitamin E (4400 IU/d) increased serum albumin fraction but did not affect different fractions of globulin (Rahmani *et al.*, 2014). Similar responses were detected in buffaloes (Helal *et al.*, 2009). In rabbits, plasma TP concentration was not significantly affected by vitamin E deficiency, but albumin levels were lower and globulin levels were higher (Diehl and Delincee, 1986). In the current study, a significant increase in serum albumin concentrations was observed for lambs received VE and VA+E treatments compared to those received VA alone (Tables, 3). This response may be physiologically useful in controlling the osmotic pressure and flow of water between blood and tissue fluids (Kobeisy *et al.*, 1997). Furthermore, the significant increases in globulin and immunoglobulin G concentrations as well as lymphocytes % of lambs treated with VA and/or VE (Tables, 2) may support the findings that both vitamins could enhance the animal 'immune function (Smith and Hays, 1987; Shinde *et al.*, 2007; Yang *et al.*, 2010).

Data also show that the other serum metabolite concentrations such as glucose, cholesterol and aspartate transaminase (AST) activity were not significantly differed due to treatments with VA, VE and VA+E (Tables, 3). These results are in agreement with other reports showed no significant changes in serum cholesterol, triglyceride concentrations or the sum of the two lipid fractions in sheep fed vitamin E alone (Njeru *et al.*, 1994). Over-supplementation of vitamin A had no effect on serum TP, albumin and globulin in dairy calves (Moosavian *et al.*, 2010). Similar findings were obtained with feeding vitamin E that did not affect serum glucose, cholesterol and AST concentrations in rats (Jang *et al.*, 1999).

Serum immune and antioxidant status:

The results showed that lambs received VA, VE and VA+E treatments had higher ($P<0.05$) serum immunoglobulin G (IgG) concentrations by 17.0, 34.4 and 22.3 %, respectively than the control (Table, 4). Lambs received VE treatment had higher ($P<0.05$) serum IgG concentrations by 15.0 % than those received VA treatment. Serum IgG concentration of lambs received the combined treatment of VA+E was not significantly differed compared with each of VA or VE alone. Earlier reports reviewed the essential role of vitamin A in immune function. According to **Smith and Hays (1987)**, with respect to immunity, supplement of vitamin A enhanced the delay of hypersensitivity, cytotoxic activity, and graft-versus-host responses to antigens. In addition, serum Igs responses and plaque-forming cell numbers were increased by vitamin A supplement. Immune enhancement accompanied by elevated T-lymphocyte numbers and interleukin-2 production. Conversely, vitamin A deficiency was associated with depressed immunity. Delayed hypersensitivity to a contact allergen, natural killer cell activity, and mitogen responses were depressed during vitamin A deficiency. Similarly, decreased serum antibody responses occurred in animals with reduced serum vitamin A concentrations. In goats, the improved antioxidant status together with the enhanced immune function, by vitamin A supplementation, indicated that vitamin A might serve as an antioxidant to protect the immune cells against oxidant stressors and

thereby maintain optimum immune function (**Yang et al., 2010**). So, the observed significant increases in serum IgG concentrations and lymphocytes percentage of lambs treated with VA in the present study (Tables 4 and 2) suggest that VA supplementation could enhance the immune response of lambs. On the other hand, VE was more potent to increase serum IgG concentration especially when compared to the effect of VA (Table, 4). Vitamin E has been implicated in stimulation of serum antibody synthesis, particularly IgG antibodies (**Tengerdy, 1980**). Vitamin E at high doses had increased serum IgG concentrations in lambs (**Gentry et al., 1991**) and in ewes (**Anugu et al., 2013**). In addition, linear increase in serum IgG titers was noted with vitamin E supplement in beef cattle (**Rivera et al., 2002**). Vitamin E can increase the viscosity of phagocyte cell membranes, leading to improved phagocytosis of foreign bodies, and may also increase production of IgG (**St-Laurent et al., 1990**). Furthermore, the protective effects of vitamin E on animal health may be involved with its role on reduction of glucocorticoids, which are known to be immunosuppressive (**Golub and Gershwin, 1985**). Metabolites concentration in serum represents a buffering state for metabolic synthesis and catabolism end products (Swenson, 1984). It could also be noticed that changes in blood hematological and biochemical parameters, in the present study, were within the normal physiological range of sheep as previously documented (**Duncan and Prasse, 1986**).

Table 3: Effects of vitamin A and vitamin E administration on serum biochemical constituents of growing Ossimi lambs (mean \pm SEM).

Parameters	Treatments				SEM	Sig.
	Control	VA	VE	VA+E		
Glucose (mg/dl)	47.4	54.7	58.5	56.2	6.3	NS
Total protein (g/dl)	6.30 c	7.54 b	7.90 a	8.15 a	0.24	*
Albumin (g/dl)	3.10 b	3.05 b	3.88 a	3.85 a	0.13	*
Globulin (g/dl)	3.34 b	4.49 a	4.19 a	4.30 a	0.09	*
Cholesterol (mg/dl)	66.5	67.9	64.7	70.0	4.3	NS
AST (U/L)	99.2	111.5	101.8	104.6	8.7	NS

a,b,c means within the same row having different superscripts significantly different

(* $P<0.05$). NS = not significant

VA = Vitamin A, VE = Vitamin E, VA+E = Vitamin A plus vitamin E.

AST = aspartate transaminase.

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Table 4: Effects of vitamin A and vitamin E administration on immunoglobulin G and antioxidant enzymes activity of growing Ossimi lambs (mean ± SEM).

Parameters	Treatments				SEM	Sig.
	Control	VA	VE	VA+E		
IgG (µg/ml)	16.6 c	19.4 b	22.3 a	20.3 ab	1.3	*
TAC (mM/L)	0.620 b	0.833 a	0.777 a	0.755 a	0.043	*
SOD (U/ml)	230.0	236.7	231.9	240.0	4.60	NS
GSH-Px (mU/ml)	4.70 c	6.71 b	8.25 a	7.44 ab	0.618	*

a,b,c means within the same row having different superscripts significantly different (* P<0.05). NS = not significant

VA = Vitamin A, VE = Vitamin E, VA+E = Vitamin A plus vitamin E.

IgG= Immunoglobulin G, TAC= Total antioxidant capacity, SOD= Superoxide dismutase,

GSH-Px = Glutathione peroxidase

Data showed a significant (P<0.05) increase in serum total antioxidant capacity (TAC) (P<0.05) by 34.4, 25.3 and 21.8 % for lambs received VA, VE and VA+E treatments vs. control, respectively (Table, 4). No significant differences in serum superoxide dismutase (SOD) activity noticed among the experimental treatment groups. There were significant (P<0.05) increases in serum glutathione peroxidase (GSH-Px) activity estimated by 42.8, 75.5 and 58.3 % for lambs received VA, VE and VA+E treatments vs. control respectively. Lambs received VE treatment had higher (P<0.05) serum GSH-Px (8.25 mU/ml) activity by 23.0 % than those received VA treatment (6.71 mU/ml). Studies focused that effect of vitamins E and A on serum antioxidant enzymes activities in sheep is limited. The present results agree with some reports on the antioxidant effects of vitamin A by **Burton and Ingold (1984)** and **Yang et al. (2010)**. In goats, dietary supplementation of vitamin A at level 2000-3000 IU/kg DM has been shown to increase (P<0.05) serum TAC and GSH-Px activities. However, the higher level of vitamin A at 5000 IU/kg DM had no effect on those enzyme activities in serum as reported by **Yang et al. (2010)**. They also found that, regardless of level of supplementation, vitamin A had no effect on serum SOD activity. The enhancing effect of vitamin A on increasing serum GSH-Px activity is likely due to the oxygen scavenging and lipo-peroxyl radical quenching function of vitamin A (**Stahl et al., 1997**). Vitamin A acts as a powerful free radical scavenger and considers the most

effective naturally occurring quencher of singlet oxygen and other free radicals (**Whittaker et al., 1996; Dugas et al., 1999**). Regarding vitamin E, it has more potent effect to increase serum GSH-Px activity especially when compared to the effect of vitamin A (Table, 4). Vitamin E is a powerful antioxidant for body defense against oxidative stress (**Ibrahim et al., 1997**). The enhancing effect of vitamin E on antioxidant enzymes activities has been shown in some investigations. In a study on goats, adding vitamin E (80 IU/ kid/d) can increase serum TAC and activities of serum SOD and GSH-Px (**Hong et al., 2010**). In male buffalo calves, blood GSH-px activity but not SOD, was significantly increased in response to vitamin E supplementation (**Shinde et al., 2008**). Hepatic SOD activity was not affected by vitamin E supplement, while GSH-px activity was significantly increased in rats. Enhanced GSH-px activity with vitamin E might aid hepatic enzymes to eliminate active oxygen in organs (**Jang et al., 1999**). Vitamin E may have a controlling effect on oxidative stress through modulation IL-2 mRNA expression of SOD (**Das et al., 2012**). As vitamin E dosage is concerned, both excess of dietary vitamin E and vitamin E deficiency can significantly depress the activity of hepatic and plasma GSH-px activity (**Yang et al., 1976**). The protective effect of vitamin E on lipid peroxidation may not due to alteration of antioxidant enzyme activity but mainly mediated through its chain-breaking antioxidant enzyme activity (**Mantha et al., 1993**).

In the current study, the significant increases in serum TAC and GSH-Px activities with combined the treatment of VA+E were comparable to that of VA or VE alone (Table, 4). It could be noticed that lambs received VA+E treatment had higher ($P<0.05$) serum GSH-Px activity by 58.3 % than control and tended to be higher by 10.9 % than lambs treated with VA. This observation could display a trend of possible beneficial effects of combined VA with VE at the level of antioxidant status. In this respect, in accordance to **Watts (1991)**, vitamin E may considered synergistic to vitamin A due to its antioxidant activity, which results in a sparing effect upon vitamin A. Likewise, loss of vitamin A from liver stores is accelerated in the presence of vitamin E deficiency as a result of increased fragility of lysosomal membranes. In addition, it has been found out that dietary supplementation of vitamin A markedly lowered *in vitro* lipid peroxidation and that vitamin E supplementation along with vitamin A still further reduced the *in vitro* lipid peroxidation of the tissues, suggesting that vitamin A also might be considered as a potential antioxidant similar to vitamin E in animal nutrition (**Kartha and Krishnamurthy, 1977**).

CONCLUSION

Based on the present results, it could be concluded that vitamins A and E combination may exerted beneficial additive effect to improve ADG and physiological responses of growing lambs. Vitamin A is more effective than vitamin E in enhancing growth performance of lambs (FBW, ADG and FCE), whereas vitamin E had more potent effect that increase serum GSH-Px activity and IgG concentration thus improve their antioxidant status and immune response.

REFERENCES

Ali, A. B. T.; G. Bomboi and B. Floris (2009): Does vitamin E or vitamin E plus selenium improve reproductive performance of rams during hot weather? Ital. J. Anim. Sci. 8:743-754.

Alosilla, C.E. Jr.; L.R. McDowell; N.S. Wilkinson; C.R. Staples and W.W. Thatcher (2007): Bioavailability of vitamin A sources for cattle. J. Anim. Sci., 85: 1235-1238.

Amer, A.H. and A.M. Hashem (2008): Reproductive performance and viability of newborns buffaloes treated antepartum with viteselen and/or ultra-corn. Slov. Vet.Res., 45: 53-60.

Antunovic, Z.; M. Speranda; Z. Steiner; M. Vegara; J. Novoselec and M. Djidara (2009): Blood metabolic profile of Tsigai sheep in organic production. Krmiva, 51:207-212.

Anugu, S.; C.S. Peterson-Wolfe and G.F. Combs Jr. (2013): Effect of the vitamin E on the immune system of ewes during late pregnancy and lactation. Small Rumin. Res., 111:83-98.

Arnett, A. M.; M. E. Dikeman; C. W. Spaeth; B. J. Johnson and B. Hildabrand (2007): Effects of vitamin A supplementation in young lambs on performance, serum lipid, and longissimus muscle lipid composition. J. Anim. Sci., 85:3062-3071.

Bedo, G.; P. Santisteban and A. Aranda (1989): Retinoic acid regulates growth hormone gene expression. Nature, 339:231-234.

Bruns, N.J. and K.E. Webb Jr. (1990): Vitamin A deficiency: serum cortisol and humeral immunity in lambs. J. Anim. Sci. 68:454-459.

Burton, G.W. and K.U. Ingold (1984): *Beta-carotene: An unusual type of lipid antioxidant.* Sci. Washington, DC. 224: 569-573.

Campbell, T.W. (1995): *Avian hematology and cytology (2nd)* - Iowa State University Press Ames, pp. 144.

Chew, B. P. (1987): Vitamin A and β -carotene on host defense. J. Dairy Sci., 70: 2732.

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- Chew, B. P. (1993):** Role of carotenoids in the immune response. *J. Dairy Sci.*, 76: 2804-2811.
- Dacie, S.J. and S.M. Lewis (1991):** *Practical hematology*. 7th Ed., Churchill Livingstone, London, UK, PP.: 118-127.
- Das, T.K.; V. Mani; S.De; D. Bnerjee; A. Mukherjee; S. Polley; N. Kewalramani and H. Kaur (2012):** Effect of vitamin E supplementation on mRNA expression of superoxide dismutase and interleukin-2 in arsenic exposed goat leukocytes. *Bull Environ. Contam. Toxicol.*, 89:1133-1137.
- Debier, C. and Y. Larondelle (2005):** Vitamins A and E: metabolism, roles and transfer to offspring. *Brit. J. Nutr.* 93:153-174.
- Diehl, J.F. and H. Delincee (1986):** Vitamin E deficiency in rabbits receiving a high PUFA diet with and without a non-absorbable antioxidant II. Incorporation of ¹⁴C-labelled glycine and L-leucine into liver and plasma proteins. *Z. Ern & hrungswiss.* 25:180-188.
- Dugas, T.R.; D.W. Morel and E.H. Harrison (1999):** Dietary supplementation with β -carotene, but not with lycopene inhibits endothelial cell-mediated oxidation of low density lipoprotein. *Free Radic. Biol. Med.*, 26:1238-1244.
- Duncan, D.B. (1955):** Multiple range test and multiple F-test. *Biometrics*, 11: 1-42.
- Duncan, J.R. and K.W. Prasse (1986):** *Veterinary Laboratory Medicine*, 2nd Ed., Iowa State University Press, USA.
- Eicher, S.D.; J.L. Morill and F. Blecha (1994):** Vitamin concentration and functions of leukocytes from dairy calves supplemented with vitamin A, vitamin E and β -carotene *in vitro*. *J. Dairy Sci.* 77: 560-565.
- El-Barody, M.A.A.; Z.B.H. Rabie and F.M.R. El-Feel (1993):** Productive and reproductive responses of pregnant Egyptian buffaloes to vitamin A injection during summer. *Minia J. Agric. Res. & Dev.* 15: 717-733.
- El-Masry, K. A.; H. M. Youssef; A. M. Abdel-Samee; I. F. M. Maria and M. K. Metawally (1998).** Effects of supplemental Zn and vitamin A on some blood biochemical and immune indices related to growth performance in growing calves. First international conference on animal production and health in semi-arid areas, El-Arish, Egypt, 1-3 Sept., 130-151.
- El-Shahat, K.H. and U.M. Abdel Monem (2011):** Effects of dietary supplementation with vitamin E and /or selenium on metabolic and reproductive performance of Egyptian Baladi ewes under subtropical conditions. *World Appl. Sci. J.*, 12:1492-1499.
- Frandsen, R.D. (1986):** *Anatomy and physiology of farm animals*. 4th Ed., Lea & Febiger, Philadelphia. Pages: 370-377.
- Gallo-Torres D.C. (1980):** *Absorption, blood transport and metabolism of vitamin E*. In: Maclin L.J. (ed.): *A Comprehensive Treatise*. Marcel Dekker, New York. Pages:170-267.
- Galyean, M.L.; L.J. Perino and G.C. Duff (1999):** Interaction of cattle health-immunity and nutrition. *J. Anim. Sci.*, 77: 1120-1134.
- Gentry, P. C.; T. T. Ross; B. C. Oetting and K. D. Birch (1991):** Effects of supplemental D-a-tocopherol on pre-weaning lamb performance, serum and colostrum tocopherol levels and immunoglobulin G titers. *Sheep Res. J.* 8:95-100.
- Golub, M.S. and M.E. Gershwin (1985):** Stress-induced immunomodulation: what is it, if it is? In "Animal Stress" (G.P. Moberg, ed.). *Am. J. Physiol. Soc.*, Bethesda, Maryland.
- Hays, V.S; D.R. Gill; R.A. Smith and R.L. Ball (1987):** The effect of vitamin E supplementation on performance of newly received stocker cattle. *Anim. Sci. Res. Rep. Oklahoma Agric. Exp. Station*, 119: 198-201.
- Helal, T.S.; F.A. Ali; O. Ezzo and M.A. El-Ashry (2009):** Effect of supplementing some vitamins and selenium during the last stage of pregnancy on some reproductive aspects of Egyptian dairy buffaloes. *J. Agri. Sci.* 19: 4289-4299.

- Hidiroglou, M (1993):** Assessment of the oral administration of a high dose of retinol on vitamin E status of sheep. *Vet. Res.*, 24:477-482.
- Hogan, J.S.; W.P. Weiss and K.L. Smith (1993):** Role of vitamin E and selenium in host defense against mastitis. *J. Dairy Sci.* 76:2795-2803.
- Hogan, J.S.; W.P. Weiss; D.A. Todhunter; K.L. Smith and P.S. Schoenberger (1992):** Bovine neutrophils responses to parenteral vitamin E. *J. Dairy Sci.* 75:399-405.
- Hong, Z.; L. Hailing; M. Hui; Z. Guijie; Y., Leyan and Y. Dubing (2010):** Effect of vitamin E supplement in diet on antioxidant ability of testis in Boer goat. *Anim. Reprod. Sci.* 117:90-94.
- Ibrahim, W., U.S. Lee, C.C. Yeh, J. Szabo, G. Bruckner and C.K. Chow (1997):** Oxidative stress and antioxidant status in mouse liver: effects of dietary lipid, vitamin E and iron. *J. Nutr.* 127: 1401-1406.
- Jang, I.S.; K.R. Chase; T.S. Kang; Y.K. Kim; C.K. Kim; J.H. Hwang; D.Y. Hwang; C.B. Choi; K.K. Jung and J.S. Cho (1999):** Effects of long-term vitamin E and butylated hydroxyl-toluene supplemented diets on murine intestinal and hepatic antioxidant enzyme activities. *Asian-Aus. J. Anim. Sci.*, 6:932-938.
- Jilani, T. and M.P. Iqbal (2011):** Does vitamin E have a role in treatment and prevention of Anemia's? *Pak. J. Pharm. Sci.*, 24:237-242.
- Kartha, V.N. and S. Krishnamurthy (1977):** Antioxidant function of vitamin A. *Int. J. Vitam. Nutr. Res.*, 47:394-401.
- Kobeisy, M.A.; L.A. Salem; M. Zenhom and M. Hayder (1997):** The effect of giving ascorbic acid on some physiological and hematological parameters of suckling lambs exposed to solar radiation and exercise. *Assuit Vet. Med. J.*, 37: 120-132.
- Kumar, M. V.; Sunvold, G. D. and Scarpace, P. J. (1999):** Dietary vitamin A supplementation in rats : Suppression of leptin and induction of UCP1 mRNA. *J. Lipid Res.*, 40:824-829.
- Liebler, D.C. (1993):** The role of metabolism in the antioxidant functions of vitamin E. *Critical Rev. Toxicol.*, 23, 147-169.
- Mantha, S.V.; M. Prasad; J. Karla and K. Prasad (1993):** Antioxidant enzymes in hyper-cholesterolemia and effects of vitamin E in rabbits. *Atherosclerosis* 101:135-144.
- Meglia, G.E.; K. Holtenius; L. Petersson; P. Ohagen and K. Waller (2004):** Prediction of vitamin A, vitamin E, selenium and zinc status of periparturient dairy cows using blood sampling during the mid dry period. *Acta Veter. Scand.*, 45:119.
- Moosavian, H. R.; M. Mohri and H. A. Seifi (2010):** Effects of parenteral over-supplementation of vitamin A and iron on hematology, iron biochemistry, weight gain, and health of neonatal dairy calves. *Food and Chem. Toxicol.* 48: 1316–1320.
- Njeru, C. A.; L. R. McDowell; N. S. Wilkinson and S. N. Williams (1994):** Assessment of vitamin E nutritional status in sheep. *J. Anim. Sci.*, 72:3207-3212.
- Nockels, C. F. and R. Blair (1996):** Antioxidants improve cattle immunity following stress. *Anim. Sci. & Tech.*, 62: 59-68.
- NRC (1985):** *Nutrient Requirements of Sheep*. 6th Ed., Washington, D.C. National Academy Press. PP. 22-23.
- Pond, W.G.; D.C. Church and K.R. Pond (1995):** Basic animal nutrition and feeding. 4th Ed. Jhon Wiley & Sons. New York. USA. Pages: 223-229.
- Qureshi, Z. I.; L. A. Lodhi; H.A. Samad; N.A. Naz and M. Nawaz (2001):** Hematological profile following immunomodulation during late gestation in buffaloes (*Bubalis Bubalus*). *Pak. Vet. J.*, 21: 148-151.
- Rahmani, M.; M. Dehghan-Banadaky; R. Kamalyan; H. Malekinejad; F. Rahmani; M.H.H. Tavatori and H. Mohammadi**

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- (2014): Effect of feeding rumen-protected choline and vitamin E on serum protein fractions, total thiol molecules and total antioxidant capacity in early lactating dairy cows. *Global J. of Anim. Sci. Res.*, 4:337-344.
- Raofi, A.; F. Asadi; S.H. Mardjanmehr and R. Kazempoor (2010):** The effects of hypervitaminosis A in sheep following intramuscular administrations of vitamin A. *Food and Chem. Toxicol.* 48:193-195.
- Reddy, P.G.; J.L. Morill; H.C. Minocha; M.B. Morill; A.D. Dayton and R.A. Frey (1986):** Effect of supplemental vitamin E on the immune system of calves. *J. Dairy Sci.*, 69:164-171.
- Rivera, J. D.; G. C. Duff; M. L. Galyean; D. A. Walker and G. A. Nunnery (2002):** Effects of supplemental vitamin E on performance, health, and humoral immune response of beef cattle. *J. Anim. Sci.*, PMID 12002330.
- Rooke, J. A.; J. J. Robinson and J. R. Arthur (2004):** Effects of vitamin E and selenium on the performance and immune status of ewes and lambs. *J. Agric. Sci.*, 142: 253-262.
- Sahin, K., N. Sahin, M. Onderci, S. Yaralioglu, O. Kucuk (2001):** Protective role of supplemental vitamin E on lipid peroxidation, vitamins E, A and some mineral concentrations of broilers reared under heat stress. *Vet. Med.*, 46:140-144.
- Salinas-Chavira, J.; A.A. Arrizon; A. Barrreas; C.Z. Chen; A. Plascencia and R.A. Zinn (2014):** Evaluation of supplemental vitamin A and E on 56-day growth performance, dietary net energy, and plasma retinol and tocopherol concentrations in Holstein steer calves. *Prof. Anim. Sci.* 30:510-514.
- SAS (2000):** SAS/STAT Guide for personal computers, SAS Inst., Cary. N.C., USA.
- Schelling, G.T.; R.A. Roeder; M.J. Garber and W.M. Pumfrey (1995):** Bioavailability and interaction of vitamin A and vitamin E in ruminants. *J. Nutr.*, 125 (6 Suppl.): 1799S-1803S.
- Semba, R.D. (1998):** The role of vitamin A and related retinoids in immune function. *Nutr. Rev.*, 56:S38-48.
- Semba, R.D. and M.W. Bloem (2002):** The anemia of vitamin A deficiency: epidemiology and pathogenesis. *Eur. J. Clin. Nutr.* 56, 271-281.
- Shetaewi, M.M.; H.A. Daghas and T.S.A. Abd El-All (1992):** Growth performance, hematology and serum profiles of coarse-wool lambs as influenced by supplemental vitamin E. *Assuit Vet. Med. J.*, 54:64-70.
- Shinde P.L.; R.S. Dass; A.K. Garg and V.K. Chaturvedi (2007):** Immune response and plasma alpha-tocopherol and selenium status of buffalo (*Bubalus bubalis*) calves supplemented with vitamin E and selenium. *Asian-Austr. J. Anim. Sci.*, 20:1539-1545.
- Shinde, P.L.; R.S. Dass; A.K. Garg and A.K. Pattanaik (2008):** Effect of vitamin E and selenium supplementation on antioxidant status of male buffalo (*Bubalus bubalis*) calves. *J. Anim. & Feed Sci.*, 17:318-327.
- Soliman, E.B. (2005):** physiological reactions and growth performance of lambs supplemented by vitamin A with zinc under summer conditions. *Assuit Vet. Med. J.* 51: 21-38.
- Smith, S. M. and C. E. Hayes (1987):** Contrasting impairments in IgM and IgG responses of vitamin A-deficient mice. (Immunity/T lymphocytes/B lymphocytes/immunoglobulins/ retinoids). *Proc. Natl. Acad. Sci.*, 84:5878-5882.
- Stahl, W.; S. Nicolai; K. Briviba; M. Hanusch and G. Broszeit (1997):** Biological activities of natural and synthetic carotenoids: Induction of gap junctional communication and singlet oxygen quenching. *Carcin*, 18: 89-92.
- St-Claire, M.B.; M.J. Kennet and C.L. Besch-Williford (2004):** Vitamin A toxicity and vitamin E deficiency in a rabbit colony.
- St-Laurent, A.; M. Hidiroglou; M. Snoddon and J.W.G. Nicholson (1990):** Response to

- dietary vitamin E in the dairy cow and its effect spontaneous oxidized flavor in milk. *Can. J. Anim. Sci.* 70:561-570.
- Swenson, M.J. (1984):** *Duke's physiology of domestic animals.* 10th Ed. Part.1. P.15, Cornell Univ. Press. Pages: 156-160.
- Tengerdy, R.P. (1980):** *Disease resistance: Immune response.* In "Vitamin E: A Comprehensive Treatise" (L.J. Machlin, ed.) Marcel Dekker, New York.
- Watts, D. L. (1991):** The nutritional relationships of vitamin A. *J. Orthomolec. Med.;* 6:27-30.
- Whittaker, P., W.G. Wamar, R.F. Chanderbhan and V.C. Dunkel (1996):** Effects of alpha-tocopherol and β -carotene on hepatic lipid peroxidation and blood lipids in rats with dietary iron overload. *Nutr. Cancer,* 25:119-128.
- Yang, N.Y.; I.B. Macdonald; I.D. Desai and M. Lee (1976):** Vitamin E supplementation and glutathione peroxidase activity. *Proc. Soc. Exp. Biol. Med.,* 151:770-774.
- Yang, W.; P. Wang; Y. Jing; Z. Yang; C. Zhang; S. Jiang and G. Zhang (2010):** Effects of vitamin A on growth performance, antioxidant status and blood constituents in lactating Grey Goat. *Amer. J. of Anim. and Vet. Sci.,* 5 : 274-281.
- Zhao, T.; H. Luo; Y. Zhang; K. Liu; H. Jia; Y. Chang; L. Jiao and W. Gao (2013)** Effect of vitamin E supplementation on growth performance, carcass characteristics and intramuscular fatty acid composition of *Longissimus dorsi* muscle in 'Tan' sheep. *Chilean, J. Agric. Res.,* 73: 358-365.

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الملخص العربي

التغيرات في الأداء الإنتاجي، المؤشرات الهيماتولوجية والبيوكيميائية للدم وحالة المناعة ومضادات الأكسدة في حملان الأوسيمي النامية نتيجة إعطاء فيتامين هـ وفيتامين أ

عصام بسيوني سليمان

قسم الإنتاج الحيواني- كلية الزراعة- جامعة المنيا

أستخدم في هذه الدراسة عدد ٣٢ حمل أوسيمي نامية بمتوسط وزن 19.08 ± 1.09 كجم وثلاثة شهور ونصف من العمر وذلك بهدف تقييم تأثير إعطاء فيتامين هـ وفيتامين أ على الأداء الإنتاجي، المؤشرات الهيماتولوجية والبيوكيميائية للدم، حالة المناعة ومضادات الأكسدة. قسمت الحيوانات عشوائياً إلى أربعة مجموعات متساوية (٨ حملان في كل منها). المجموعة الأولى للمقارنة (كنترول)، المجموعة الثانية (VA) أعطيت فيتامين أ بمعدل ٥٠.٠٠٠ وحدة دولية/رأس/كل أسبوعين، والمجموعة الثالثة (VE) أعطيت فيتامين هـ بمعدل 400 ملجم/رأس/كل أسبوعين، والمجموعة الرابعة (VA+E) أعطيت فيتامين أ بمعدل ٥٠.٠٠٠ وحدة دولية + فيتامين هـ بمعدل 400 ملجم/رأس/كل أسبوعين.

وقد أظهرت النتائج ما يلي :- سجلت حملان المعاملات (VA)، (VA+E) قيم أعلى معنوياً لمتوسطات وزن الجسم النهائي مقارنة بحملان الكنترول والمعاملة (VE). زادت متوسطات معدل الزيادة اليومية في الوزن معنوياً لحملان المعاملة (VA)، (VA+E) مقارنة بالكنترول وحملان المعاملة (VE). كما كان هناك زيادة معنوية في متوسطات معدل الزيادة اليومية في الوزن لحملان المعاملة المشتركة (VA+E) عند مقارنتها بحملان كل من المعاملة (VE) والمعاملة (VA) منفردتين. لم تلاحظ أية تغييرات معنوية في إجمالي استهلاك المادة الجافة لحملان المعاملات مقارنة بالكنترول، بينما حدث تحسین معنوی (P < 0.05) في معدلات تحويل الغذاء لحملان المعاملات (VA)، (VA+E) مقارنة بحملان المعاملة (VE) والكنترول.

- أظهرت حملان المعاملات (VA)، (VE)، (VA+E) زيادة معنوية في تركيز هيوجلوبين الدم، بينما زاد تركيز الهيموجلوبين وعدد كرات الدم الحمراء والمكونات الخلوية للدم لحملان المعاملات (VE)، (VA+E) مقارنة بحملان الكنترول. لم تلاحظ أية تغييرات معنوية في العدد الكلي لكريات الدم البيضاء وكذلك نسبة الكريات حمضية الصبغ، والكريات قاعدية الصبغ والكريات الأحادية نتيجة للمعاملات بينما أظهرت حملان المعاملات (VA)، (VE)، (VA+E) زيادة معنوية في نسبة الكريات الليمفاوية مقارنة بالكنترول. أظهرت حملان المعاملة (VE) إنخفاض معنوی في نسبة الكريات المتعادلة مقارنة بحملان الكنترول والمعاملات (VA)، (VA+E).

- أظهرت حملان المعاملات (VA)، (VE)، (VA+E) زيادة معنوية في تركيزات السيرم من البروتين الكلي والجلوبولين مقارنة بالكنترول، بينما زادت معنوياً تركيزات السيرم من البروتين الكلي والألبومين لحملان المعاملة المشتركة (VA+VE) مقارنة بحملان المعاملة (VA). أظهرت حملان المعاملات (VA)، (VE)، (VA+E) زيادة معنوية في تركيزات السيرم من الإمينوجلوبولين (IgG) مقارنة بالكنترول فيما زاد تركيز السيرم من الإمينوجلوبولين (IgG) في حملان المعاملة (VE) مقارنة بحملان المعاملة (VA). كان هناك زيادة معنوية في إجمالي القدرة المضادة للأكسدة (TAC) ونشاط إنزيم الجلوتاثيون بيروكسيداز (GSH-Px) في السيرم لحملان المعاملات (VA)، (VE)، (VA+E) مقارنة بالكنترول، فيما زاد نشاط إنزيم (GSH-Px) في السيرم معنوياً لحملان المعاملة (VE) مقارنة بحملان المعاملة (VA). لم تلاحظ أية تغييرات معنوية في تركيزات سيرم الدم من الجلوكوز، الكوليستيرول وكذلك في مستويات نشاط إنزيم (AST) ونشاط إنزيم (SOD) في حملان المعاملات (VA)، (VE)، (VA+E) مقارنة بالكنترول.

بناءً على النتائج المقدمة، فإن إعطاء فيتامين هـ وفيتامين أ للحملان النامية يمكن أن يؤدي إلى تأثيرات مفيدة مشتركة لهذه الفيتامينات لتحسين معدل الزيادة اليومية في الوزن وكذلك الإستجابات الفسيولوجية لهذه الحملان. وقد كان فيتامين أ أكثر فاعلية لتعزيز أداء النمو بينما كان لفيتامين هـ تأثير أكثر فاعلية لتعزيز الإستجابة المناعية وحالة مضادات الأكسدة.