

## PRODUCTIVE PERFORMANCE AND SOME BIOCHEMICAL INDICES OF OSSIMI EWES AND THEIR LAMBS TO DIETARY SELENIUM SUPPLEMENTATION

E. M. Ibrahim<sup>1\*</sup> and M.Y. Mohamed<sup>2</sup>

<sup>1</sup> Anim. Prod. Depart., Fac. of Agric., Minia Univ., Egypt.

<sup>2</sup> Anim. Prod. Res. Institute, Agric. Res. Cent., Ministry of Agric., Egypt.

\*Corresponding author E-mail: [emad\\_i@mu.edu.eg](mailto:emad_i@mu.edu.eg)

### ABSTRACT

A total number of 24 Ossimi pregnant ewes were used to evaluate the effects of inorganic selenium (Se) and Nano-Se supplementation upon productive performance, nutrients digestibility, some biochemical indices and antioxidant status of Ossimi ewes and their suckling lambs. The ewes were fed on a basal diet (control) supplemented with Se at 0.3 mg as sodium selenite (T1) or Nano-Se (T2) /kg DM. The results showed that OM, CP, EE, CF and ADF digestibility were increased ( $P<0.05$ ) for ewes fed T2 compared to those fed T1 or the control. Birth weight, FBW and ADG were improved ( $P<0.05$ ) for lambs born to ewes fed T1 or T2 diets vs. control. Milk yield, FCM, milk fat yield and milk protein yield recorded higher ( $P<0.05$ ) values for ewes fed T1 or T2 than the control one. Serum IgG, total protein, albumin, globulin, thyroid hormones, total antioxidant capacity and glutathione-S-transferase levels were higher ( $P<0.05$ ) for both ewes fed T1 or T2 and their lambs than the respective control. Serum cholesterol, liver enzymes (ALT and AST) and urea concentrations showed a decrease ( $P<0.05$ ) for ewes fed T1 or T2 and their respective lambs compared to those fed the control one.

It is concluded that dietary supplementation of Nano-Se at 0.3 mg/Kg DM to Ossimi ewes, at late gestation and the suckling period, was more potent than inorganic Se to improve their nutrients digestibility, productive performance, some serum biochemical indices and antioxidant status, leading to better growth performance and health of their suckling lambs.

**Keywords:** *Inorganic Selenium, Nano-selenium, digestibility, productive performance, serum biochemical indices, Ossimi ewes.*

### INTRODUCTION

Nutrients with antioxidants play crucial roles in immunity function and growth promoters, and their addition to feed could improve the antioxidant performance of ruminants (Tian *et al.*, 2018; Tian *et al.*, 2022). Selenium (Se) is among these nutrients which, it is a component of selenoenzyme glutathione peroxidase (GSH-Px), metabolizing hydrogen peroxide and lipid peroxide, thus neutralizing free radicals and

promoting immunity functions (Silveira *et al.*, 2021; Salles *et al.*, 2022). Hence, dietary Se supplementation could affect the ruminal fluid microbial function via its antioxidant effects and the modulation of the immune function (Hendawy *et al.*, 2022; Tian *et al.*, 2022). As well, Se is a key structural component in selenocysteine, that exists in the active sites of iodothyronine deiodinase and thioredoxin reductases enzymes that are involved in thyroid hormones metabolism (Adadi *et al.*, 2019; Arshad *et al.*, 2021). So, it is believed



that Se can influence the growth of ruminants through its regulation of thyroid hormone levels (Kieliszek and Blazejak, 2016).

It has been reviewed that Se levels in soils and plants are low in many regions worldwide, including Egypt (Abdelrazek and Fayed 2025), leading to wide spread of Se deficiency symptoms in ruminants (Wang *et al.*, 2019; Rajab *et al.*, 2023). The Se deficiency in ruminants could induce metabolic disorders, reduced productivity, health issues and even mortality resulting in financial losses for sheep producers (Hofstee *et al.*, 2020). In ewes, a specific clinical sign of Se deficiency is muscular dystrophy, particularly affecting lambs born to selenium-deficient mothers (Lee *et al.*, 2019). Additionally, Se deficiency contributes to fertility issues, abortions, retained placenta, and health problems in newborns, such as higher neonatal mortality, reduced vitality, impaired suckling reflex, and weakened immune function, making the lambs more vulnerable to infections (Milewski *et al.*, 2021; Pecoraro *et al.*, 2022). During the third trimester of pregnancy period, fetal growth accelerates, leading to an increased transfer of maternal Se to the developing fetus. Consequently, insufficient Se intake in pregnant and lactating ewes may induce Se deficiency in nursing ewes and their newborns lambs (Goof 2018; Lee *et al.*, 2019), potentially impacting their immunity and productive performance.

Nanotechnology offers an innovative approach to enhance animal health and productive performance via providing nutrients and improving the quality of animal-derived products (Malyugina *et al.*, 2021). Nanoparticles, with their unique physicochemical properties such as small size (1–100 nm), high stability, hydrophobicity, large surface area, enable efficient dispersion, cellular uptake, and interaction with biological systems (Malyugina *et al.*, 2021; Gu and Gao,

2022). Selenium nanoparticles, in particular, have gained attention for their high bioavailability, low toxicity, and superior efficacy in enhancing growth, reproduction, immunity, antioxidant enzyme activity and nutrient digestibility in ruminants, making them a sustainable and effective alternative for dietary supplementation of traditional inorganic and organic Se sources (Malyugina *et al.*, 2021; Han *et al.*, 2021; Ferrari *et al.*, 2023). Therefore, the present study aimed to evaluate the effects of inorganic Se and Nano-Se upon productive performance, nutrients digestibility and some biochemical indices as well as the antioxidant status of Ossimi ewes and their suckling lambs.

## MATERIALS AND METHODS

### Animals:

Twenty-four Ossimi pregnant ewes averaged  $38.06 \pm 2.09$  kg and 2.5-3 years old at 4 -6 weeks of late-gestation were used in this study, which was carried out at the Farm of Animal Production Department., Faculty of Agriculture, Minia University, El-Minia, Egypt.

### Feeding and Management:

Ewes were fed on a concentrate feed mixture (CFM) to cover their nutrients requirements according to their live body weight (NRC, 2007). The ewes were randomly divided into three equal groups (8 ewes each) of similar initial body weights. The 1<sup>st</sup> group was fed on un-supplemented basal diet (control), but the 2<sup>nd</sup> and 3<sup>rd</sup> groups were fed on the basal diet supplemented with 0.3 mg Se as sodium selenite (T1) or Nano-Se (T2) /kg DM, respectively. The basal diet was consisted of CFM (40 % wheat bran, 30 % yellow corn, 17 % soybean meal, 10 % wheat straw, 2 % calcium carbonate and 1 % sodium chloride) and alfalfa hay (AH) that offered *ad libitum*.



The ewes were housed inside window stables and fed by using group feeding system. The calculated concentration of Se in the CFM was 0.19 mg/kg DM. The NRC (2007) have been recommended a dietary level of 0.30 mg Se / kg DM for sheep. The ewes were fed on supplemented diet treatments starting at 4 – 6 weeks of late-gestation and during the suckling period for three months. Feed was daily offered twice at 8 am and 2 pm and fresh water was always available to the ewes. The mean DMI, at the last week of each month, was considered in calculation of digestibility and feeding values of dietary treatments. The body weights of ewes in different experimental groups were recorded at the beginning of experiment during late-gestation and at lambing and then at every month during suckling period. Body weights of lambs born to ewes in each group were recorded within 24 hours from birth and then biweekly during suckling period for the consecutive three months and the averages of lambs' daily weight gain were calculated.

### Dietary Sampling and laboratory analysis:

Dietary samples were daily collected at the last week of each month during the experimental period, and a composite sample was performed. A portion of the composite sample was dried at 105 °C in a forced air oven till constant weight for DM determination. The rest of the composite sample was dried at 70 °C for a constant weight, ground and kept in closely tied jars for laboratory analysis. Diets were analyzed for dry matter (DM), organic matter (OM), crude protein (CP), crude fiber (CF), ether extract (EE) and ash according to AOAC (2019). Neutral detergent fiber (NDF) and acid detergent fiber (ADF) were determined according to Goring and Van Soest (1970). Daily grab fecal samples of each month were collected before feeding at 7 am and 2 pm for each ewe at the last week of each month and mixed, dried at 70 °C till constant weight and analyzed for DM, OM, CP, CF, NDF, ADF, EE and ash. Total tract digestibility of DM, OM, CP, CF, NDF, ADF, EE and NFE were determined using acid insoluble ash as an internal marker according to Van Keulen and Young (1977). Approximate analysis of CFM, AH and total mixed ration (TMR) are presented in Table (1).

**Table (1): Proximate analysis of concentrate feed mixture, alfalfa hay and total mixed ration fed to Ossimi ewes (% on DM basis).**

Item	CFM	AH	TMR
DM	88.92	84.20	87.27
OM	92.24	88.51	90.93
CP	17.88	14.15	16.58
EE	3.45	1.04	2.61
CF	10.58	24.98	15.62
NFE	60.32	48.33	56.12
NDF	26.26	60.20	38.14
ADF	13.89	48.34	25.95
Ash	7.76	11.49	9.07

CFM: Concentrate feed mixture contained 40 % wheat bran, 30 % yellow corn, 17 % soybean meal, 10 % wheat straw, 2 % calcium carbonate and 1 % sodium chloride. AH: Alfalfa hay. TMR: Total mixed ration



### Milk sampling and analysis:

Daily milk yield was measured biweekly from each ewe starting from the fifth day of lambing till weaning. Milk yield was determined using lamb-sucking technique according to Ashmawy (1980). Lambs were separated from their mothers at 4.0 pm on the day before measuring milk yield. In the following day, lambs were weighed at 8.0 am, and left to suckle their dams till satisfied, then reweighed and kept away from their mothers. The residual milk in the udder of each dam was hand-milked and weighed. At 4.0 pm, the lambs were weighed again before and after suckling and the residual milk in the udder was also hand-milked and weighed. The amount of milk consumed by each lamb in the morning and afternoon was calculated by the differences between the weight recorded before and after suckling. The ewes' hand-milked yield, in the morning and afternoon, was added to the daily milk intake by her suckling lambs to give an estimate of 24-hr milk production. Biweekly, representative milk samples were individually collected post-lambing till weaning. Milk samples were analyzed for protein, fat, lactose and solid not fat (SNF) using infrared spectrophotometry MilkoScan (model 130 series\_ type 10900 FOSS electric - Denmark). Fat corrected milk yield (FCM) was calculated by the equation: **(FCM (6%) for milk sheep) = daily milk yield  $\times$  (0.428 + 0.095  $\times$  fat %)** according to Economides and Louca (1981).

### Serum bio-chemical indices analysis:

Non-heparinized blood samples were collected from the jugular vein of the experimental ewes and their suckling lambs every two weeks before feeding and drinking. Blood samples were left to clot at room temperature for at least 4 h, then the clots were removed and sera were cleared by centrifugation at 1500 $\times$ g for 20 min and stored

at -20 °C for later assay. Serum total protein, albumin, glucose, cholesterol, aspartate transaminase (AST), alanine aminotransferase (ALT) and urea were determined colorimetrically using Bio-diagnostic product kits (Egypt). Serum globulin concentrations were calculated by difference between total protein and albumin concentrations. Levels of immunoglobulin G (IgG) were quantified three times at 12-, 24- and 48-hours post-lambing using bovine IgG ELISA kits (Alpha Diagnostic international, Texas, USA and Kamiya Biomedical Company, Seattle, Washington, USA, respectively). Serum triiodothyronine (T<sub>3</sub>) and thyroxine (T<sub>4</sub>) concentrations were determined by ELISA (Enzyme-Linked Immune-Sorbent Assay) technique using EIA kits (Prechek Bio, Inc., Atlaslink technology, California-USA). The T<sub>3</sub> / T<sub>4</sub> ratio was calculated. Serum total antioxidant capacity (TAC) and glutathione-S-transferase (GST) levels were analyzed colorimetrically by STAT-LAB SZSL60-SPECTRUM, using Bio-diagnostic kits (Bio-diagnostic Company, Egypt).

### Statistical analysis:

Data were analyzed by least square means analysis of variance using General Linear Model procedure of the statistical analysis system (SAS, 2012). The model used to analyze the different traits studied for ewes or lambs was as follows:

$$Y_{ij} = \mu + T_i + e_{ij}$$

Where: Y<sub>ij</sub>: observation,  $\mu$ : General mean; T<sub>i</sub>: Effect of i<sup>th</sup> treatments and e<sub>ij</sub>: random error. Duncan's Multiple Range test was used to detect differences between means of the experimental groups (Duncan, 1955).



## RESULTS AND DISCUSSION

### Nutrients digestibility and feeding values of the experimental diets:

The results presented in Table (2) have shown the effect of inorganic Se (Sodium selenite, T1) or Nano-Se (T2) dietary supplementation on nutrients digestibility and nutritive values of the experimental diets. The data revealed that DM and NDF digestibility were increased ( $P < 0.05$ ) for ewes fed T1 or T2 compared to that of control ration. Meanwhile, values of OM, CP, EE, CF and ADF digestibility were greater ( $P < 0.05$ ) for ewes fed T2 than those fed T1 or control with significant ( $P < 0.05$ ) differences among ewes fed T1 and T2. As far as nutritive values are concerned, values of DCP were increased ( $P < 0.05$ ) for ewes fed T2 compared to T1 or control with a significant ( $P < 0.05$ ) difference among ewes fed T1 and T2. Meanwhile, values of TDN were higher ( $P < 0.05$ ) for ewes fed T1 or T2 compared to control with insignificant differences between the two supplements. These findings related to nutrients digestibility and nutritive values are in agreement with other studies (Zhang *et al.*, 2020 and Pan *et al.*, 2021), confirming that Se supplementation can improve nutrients digestibility in ruminants, and this may be ascribed to the increase in rumen microbial counts and enzyme activity (Liu *et al.*, 2019; Cui *et al.*, 2021). In the same way, when Se was added at 0.3 mg to the basal diet in any form (inorganic, organic and Nano) resulted in a significant ( $P < 0.05$ ) increase in the digestion coefficients of DM, OM, CP, EE, CF, NDF, ADF and NFE compared to those of control (Vajpeyee *et al.*, 2024). In another study, the digestibility of DM, OM, CP, NDF and ADF were also increased ( $p < 0.05$ ) with Nano-Se supplemented diet in dairy cows, suggesting that adding Nano-Se at a moderate dose (0.3 mg Se /kg DM) was beneficial for nutrients digestion (Liu *et al.*, 2024). Taken

together, dietary Se supplementation, in ruminants, can positively impacted the fermentation in multiple ways: modifying the microbial structure of the rumen promoting the growth of bacteria that obtain more selenium, digest fiber better, having probiotic/antioxidant effects; producing more VFA, favorable pH levels can be achieved, rumen resistance to pathogens can be increased, inflammatory and oxidative stress signaling cascades can be modulated; and finally Se as an antioxidant/anti-inflammatory, along with production of VFA and promoting beneficial bacteria growth, may be corrected the oxidative stress and inflammation in the digestive tract, leading to optimal gastrointestinal functioning (Hendawy *et al.*, 2022). This ongoing discussion could explain the improvement in nutrients digestibility and feeding values that were observed in the present study, especially in response to dietary Nano-Se supplementation.

### Productive performance of ewes and their suckling lambs:

The results presented in Table (3) indicated that there were no significant differences in IBW, FBW and DMI among ewes fed T1 (Sodium selenite), T2 (Nano-Se) or control. Meanwhile, values of DCPI and TDNI were increased ( $P < 0.05$ ) for ewes fed T1 or T2 when compared to control. Accordingly, DM, OM and CP intakes were found to be similar among the control and Se-supplemented groups (Kumar *et al.*, 2022). This may indicate that supplementation of different sources of Se at 0.3 ppm level had no effect on palatability and feed intake. In the same way, Liu *et al.*, (2024) noticed that dietary Nano-Se supplementation in dairy cows had no effect on DMI. The present results are in the same line with previous works, which indicated that either levels or sources of Se did not influence FBW or DMI in Awassi ewes (Kasim *et al.*, 2023; Kareem *et al.*, 2024).



**Table 2 . Effects of inorganic Se or Nano-Se dietary supplementation on nutrients digestibility and nutritive values of experimental diets (Mean  $\pm$  SEM).**

Parameters	Treatments			± SEM
	Control	T1	T2	
Nutrients digestibility (%):				
DM	77.47 <sup>b</sup>	80.38 <sup>a</sup>	81.11 <sup>a</sup>	0.30
OM	78.98 <sup>c</sup>	80.78 <sup>b</sup>	81.69 <sup>a</sup>	0.17
CP	70.21 <sup>c</sup>	73.49 <sup>b</sup>	78.44 <sup>a</sup>	0.16
EE	71.37 <sup>b</sup>	72.42 <sup>b</sup>	76.09 <sup>a</sup>	0.21
CF	68.07 <sup>c</sup>	70.59 <sup>b</sup>	73.94 <sup>a</sup>	0.15
NDF	54.24 <sup>b</sup>	56.57 <sup>a</sup>	57.73 <sup>a</sup>	0.52
ADF	42.47 <sup>c</sup>	44.79 <sup>b</sup>	50.24 <sup>a</sup>	0.32
Nutritive values (%):				
DCP	11.64 <sup>c</sup>	12.18 <sup>b</sup>	13.01 <sup>a</sup>	0.03
TDN	67.51 <sup>b</sup>	70.41 <sup>a</sup>	71.55 <sup>a</sup>	0.36

a,b,c means within the same row having different superscripts significantly different ( $P < 0.05$ ).

T1: Inorganic Se (0.3 mg/kg DM), T2: Nano-Se (0.3 mg/kg DM).

In case of lambs, Table (3) showed that values of BW, FBW and ADG were improved significantly ( $P < 0.05$ ) by 14.25 and 31.84; 10.05 and 25.32; 8.97 and 23.74 % for lambs born to ewes fed T1 or T2 diets vs. control, respectively. Also, the differences in BW, FBW and ADG between lambs born to ewes fed T1 and T2 was significant ( $P < 0.05$ ) in favor of T2. This improvement may be explained as Se may affect growth performance via activating the conversion of tetraiodothyronine ( $T_4$ ) into the more active triiodothyronine ( $T_3$ ), a process mediated by the enzyme 5-iodothyronine deiodinase, which is a Se-dependent selenoprotein, and it is also the last selenoprotein whose activity is affected in the case of Se deficiency (Li *et al.*, 2022). Furthermore, the improvement in growth performance could be ascribed to the involvement of Se in regulating several

enzymatic systems that interfere with energetic metabolism (Saleh and Ebeid, 2019). Besides, sodium selenite and Nano-Se were shown to increase serum  $T_3$  concentration and participated in modulating mRNA expression of IGF-I and insulin receptors, thus consequently promoting growth performance traits (Aghwan *et al.* 2013). As well, Se is transmitted through the placenta to the fetus, milk and colostrum to the newborn, resulting in improved immunity and growth (Chen *et al.*, 2019). These findings are supported by previous studies (Yaghmaie *et al.* 2017; Ibrahim and Mohamed, 2018; Salam *et al.*, 2021; Vajpeyee *et al.*, 2024). On the other hand, some studies showed that Nano-Se supplementation had no significant effect on growth performance of Moghani lambs (Ghaderzadeh, 2016) or Barbari bucks (Kumar *et al.*, 2022).



**Table 3: Effects of inorganic Se or Nano-Se dietary supplementation on growth performance of ewes and their sucking lambs (Mean  $\pm$  SEM).**

Parameters	Treatments			± SEM
	Control	T1	T2	
Ewes:				
IBW (late-gestation), kg	36.92	38.67	38.58	2.09
FBW (kg/day)	38.50	39.50	39.67	1.68
AHI (kg/day)	0.49	0.51	0.51	0.02
CFMI (kg/day)	0.91	0.94	0.94	0.04
DMI (kg/day)	1.40	1.45	1.45	0.07
DCPI (g/head/day)	162.96 <sup>c</sup>	176.61 <sup>b</sup>	188.65 <sup>a</sup>	0.58
TDNI (g/head/day)	945.14 <sup>b</sup>	1020.95 <sup>a</sup>	1037.48 <sup>a</sup>	5.32
Suckling lambs:				
Birth weight (kg)	3.58 <sup>c</sup>	4.09 <sup>b</sup>	4.72 <sup>a</sup>	0.09
FBW (kg)	17.42 <sup>c</sup>	19.17 <sup>b</sup>	21.83 <sup>a</sup>	0.39
ADG (g/day)	184.45 <sup>c</sup>	201.00 <sup>b</sup>	228.23 <sup>a</sup>	5.42

a,b,c means within the same row having different superscripts significantly different ( $P < 0.05$ ).

T1: Inorganic Se (0.3 mg/kg DM), T2: Nano-Se (0.3 mg/kg DM).

Data presented in Table (4) indicated that milk yield, FCM, fat and protein yields recorded higher ( $P < 0.05$ ) values for ewes fed T1 or T2 than those of control one. Meanwhile, feed efficiency as FCM/DMI and lactose yield were increased ( $P < 0.05$ ) for ewes fed T2 compared to those fed T1 or control. The observed positive effect of Se supplementation on milk production could be associated with the improvement that noticed in nutrients digestibility (Table 2), serum metabolites (Table 6) and antioxidant status (Table 7) of ewes in the present study. The increment that occurred in milk production could be explained in the way that Se or Nano-Se is necessary for ewes to enhance their immune functions (Tian *et al.*, 2022), acting as a scavenger of oxygen-free radicals (Silveira *et al.*, 2021) and improving intestinal absorption (Gangadoo *et al.*, 2020) reflecting on optimizing milk production in small ruminants (Hashem and EL –Zarkouny, 2016). In addition, this improvement may be associated with improved fiber digestion (Table, 2) because Se supplementation improves the activity of

fibrolytic bacteria in the rumen and absorption in the intestine (Rabee *et al.*, 2023). As well, this improvement in milk production following Se supplementation could result from increased energy availability rather than solely from reducing oxidative stress or sustaining redox balance. This is because generating antioxidant defenses and repairing cellular damage caused by free radicals demand significant energy expenditure. By ensuring adequate Se levels in plasma to stabilize redox homeostasis, this energy can be preserved and redirected toward productive processes such as milk synthesis. Furthermore, the increase in milk yield might also stem from reduced free radical damage in mammary gland tissue, enhancing the efficiency of milk synthesis by supporting healthier gland function (Arshad *et al.*, 2021). These results are reinforced with previous reports which indicated that either levels or sources of supplemental Se increased milk yield, milk fat yield and milk protein yield in dairy cows (Ullah *et al.*, 2019; Zhang *et al.*, 2020; Liu *et al.*, 2024); Zaraibi goats (Gaafar *et al.*, 2021).



**Table 4: Effects of inorganic Se or Nano-Se dietary supplementation on milk production and its composition of Ossimi ewes.**

Parameters	Treatments			±SEM
	Control	T1	T2	
<b>Milk yield (g/head/day)</b>	<b>359.79<sup>c</sup></b>	<b>405.67<sup>b</sup></b>	<b>474.50<sup>a</sup></b>	<b>4.47</b>
<b>FCM (g/head/day)</b>	<b>361.76<sup>c</sup></b>	<b>408.52<sup>b</sup></b>	<b>480.43<sup>a</sup></b>	<b>5.27</b>
<b>DMI (kg/day)</b>	<b>1.40</b>	<b>1.45</b>	<b>1.45</b>	<b>0.07</b>
<b>FCM/DMI</b>	<b>0.26<sup>b</sup></b>	<b>0.28<sup>b</sup></b>	<b>0.33<sup>a</sup></b>	<b>0.01</b>
<b>Milk composition (g/kg)</b>				
<b>Fat</b>	<b>6.08</b>	<b>6.09</b>	<b>6.15</b>	<b>0.10</b>
<b>Protein</b>	<b>4.31</b>	<b>4.68</b>	<b>4.74</b>	<b>0.14</b>
<b>Lactose</b>	<b>4.11</b>	<b>4.14</b>	<b>4.18</b>	<b>0.20</b>
<b>Solid not fat</b>	<b>9.14</b>	<b>9.55</b>	<b>9.71</b>	<b>0.24</b>
<b>Ash</b>	<b>0.72</b>	<b>0.74</b>	<b>0.79</b>	<b>0.03</b>
<b>Total solids</b>	<b>15.22</b>	<b>15.66</b>	<b>15.87</b>	<b>0.28</b>
<b>Milk constituents yield (g/d)</b>				
<b>Fat</b>	<b>21.87<sup>c</sup></b>	<b>24.73<sup>b</sup></b>	<b>29.19<sup>a</sup></b>	<b>0.45</b>
<b>Protein</b>	<b>15.50<sup>c</sup></b>	<b>19.00<sup>b</sup></b>	<b>22.51<sup>a</sup></b>	<b>0.56</b>
<b>Lactose</b>	<b>14.77<sup>b</sup></b>	<b>16.81<sup>b</sup></b>	<b>19.81<sup>a</sup></b>	<b>0.76</b>

a,b,c means within the same row having different superscripts significantly different (P<0.05).

T1: Inorganic Se (0.3 mg/kg DM), T2: Nano-Se (0.3 mg/kg DM).

As shown in Table (4), no significant changes were noticed in milk composition (g/kg) among ewes fed T1 or T2 when compared to control. In this regard, some studies have shown that Se supplementation had no effect on milk components (Bagnicka *et al.*, 2017; Vasil *et al.*, 2017; Han *et al.*, 2021; Barcelos *et al.*, 2023; Liu *et al.*, 2024).

### **Serum biochemical indices of ewes and their suckling lambs:**

The data presented in Table (5) showed the concentrations of serum IgG (µg/ml) for Ossimi ewes and their lambs at 12, 24 and 48 hrs. post lambing. The results indicated that serum IgG concentrations for ewes fed T1 (Sodium selenite) or T2 (Nano-Se) and their respective lambs were increased (P<0.05) at 12, 24 and 48 hrs. post-lambing compared to

those of control diet. It was clear to notice that serum IgG concentrations of the ewes fed T2 and their suckling lambs were higher (P<0.05) than those fed T1 or control at all examined times post-lambing. These findings could be explained as Se is crucial for modulating immune function, in particular non-specific immune response and its deficiency has been associated with impaired immune system performance (Milewski *et al.*, 2021; Pecoraro *et al.*, 2022; Tian *et al.*, 2022). According to Hall *et al.*, (2013), Se supplementation may improve the lymphocytes proliferation, which may signify the increase in IgG concentration in the present study. Similar results were documented with Nano-Se supplementation in suckling lambs (Salam *et al.*, 2021) and in suckling Friesian calves (Shams *et al.*, 2020).



**Table 5 : Effects of inorganic Se or Nano-Se dietary supplementation on serum immunoglobulin G (IgG, mg/ml) concentrations of Ossimi ewes and their lambs (Mean  $\pm$  SEM).**

Time post lambing	Treatments			±SEM
	Control	T1	T2	
Ewes:				
12 hrs.	7.37 <sup>c</sup>	8.66 <sup>b</sup>	9.43 <sup>a</sup>	0.07
24 hrs.	8.78 <sup>c</sup>	10.07 <sup>b</sup>	11.51 <sup>a</sup>	0.15
48 hrs.	8.34 <sup>c</sup>	9.73 <sup>b</sup>	10.69 <sup>a</sup>	0.12
Suckling Lambs:				
12 hrs.	8.25 <sup>c</sup>	9.50 <sup>b</sup>	10.20 <sup>a</sup>	0.13
24 hrs.	9.77 <sup>c</sup>	11.00 <sup>b</sup>	11.91 <sup>a</sup>	0.21
48 hrs.	9.47 <sup>c</sup>	10.70 <sup>b</sup>	11.48 <sup>a</sup>	0.16

a,b,c means within the same row having different superscripts significantly different ( $P < 0.05$ ).

T1: Inorganic Se (0.3 mg/kg DM), T2: Nano-Se (0.3 mg/kg DM).

The data presented in Table (6) summarized the effects of Se supplementation on some serum biochemical indices for the ewes and their lambs. Most of the determined biochemical indicators were within the normal reference values (Kaneko *et al.*, 2008) indicating proper nutrition and normal metabolic processes. The results showed that serum concentrations of total protein, albumin and globulin were increased ( $P < 0.05$ ) for ewes fed T1 or T2 and their respective lambs compared to those fed control. In this regard, it has been suggested that Nano-Se supplementation may promote the efficiency of protein utilization, improve ruminal fermentation and nutrients digestibility (Shi *et al.*, 2011 and Wang *et al.*, 2019), thus could lead to the increment that occurred in serum metabolites observed in the presented study. In the same way, Liu *et al.*, (2024) reported that adding Nano-Se to dairy cows' diets at 0.1, 0.2 and 0.3 mg/kg DM had increased ( $P < 0.05$ ) blood metabolites concentrations compared to a control diet.

No significant changes were observed in serum glucose concentrations among ewes fed T1 or T2 and their respective lambs compared to those fed control (Table, 6). These results are supported by some works which indicated

that dietary Se supplementation exhibited no effects on serum glucose of male buffalo calves (Mudgal *et al.*, 2008); ewe lambs (Mousaie *et al.*, 2014) and dairy goats and their respective kids (Barcelos *et al.*, 2022)

As shown in Table (6), serum cholesterol concentrations were decreased ( $P < 0.05$ ) for ewes fed T1 or T2 as well as their suckling lambs compared to their respective control with no significant differences between T1 and T2. The decline in total cholesterol levels could point out the positive effect of dietary Nano-Se supplementation on energy metabolism and lipid profile (Ibrahim and Mohamed, 2018). The physiological status of animals may also affect their levels of total cholesterol, and the lower levels are related to an increased energy requirement (Halawa *et al.*, 2023a). The present results are in the same line with previous works which indicated that Nano-Se supplementation decreased serum cholesterol levels in Karadi lambs (Bolshakova and Aljaf 2022) and Dorper Sheep (Budak, 2024).

As shown in Table (6), the levels of AST and ALT showed a significant decrease ( $P < 0.05$ ) for ewes fed T1 or T2 rations, and their suckling lambs compared to their respective control. They also decreased



( $P < 0.05$ ) for ewes fed T2 and their lambs more than those fed T1. The decreased activity of these enzymes observed in this study, however such levels were within the normal values (Lepherd *et al.*, 2009) and may indicate that Se supplementation had no adverse effect on liver health status (Alahmadi *et al.*, 2020). In this regard, earlier studies showed that serum AST and ALT activities significantly increased in case of liver necrosis incidence or other diseases that damaged liver tissue (Alimohamady *et al.*, 2013; Mousaie *et al.*, 2014). Moreover, these decreases in AST and ALT activities, observed in the current study, could be attributed to the role of Se in removing hydrogen peroxide via the action of the glutathione peroxide enzyme (Haenlein and Anke 2011; Abdel Ghfar *et al.*, 2022). These results are reinforced by previous reports which indicated that Nano-Se supplementation decreased AST and ALT activities in Ossimi ewes (Halawa *et al.*, 2023a); male lambs (Aljaf and Bolshakova, 2023) and Dorper sheep (Budak, 2024).

The data in Table (6) showed that urea concentrations were decreased ( $P < 0.05$ ) for ewes fed T1 or T2 and their suckling lambs compared to their respective controls. They also decreased ( $P < 0.05$ ) for ewes fed T2 and their lambs more than those fed T1. The significant decrease in urea concentrations noticed for ewes receiving treatments, in particular Nano-Se, as compared to the control may signify an improvement in normal kidney functions. In this respect, Ucar *et al.*, (2010) found that Se, as an antioxidant, helps the body to get rid of urea damaging effects. This could account for the decreased urea levels observed in the presented study, suggesting a protective role of Nano-Se against kidney dysfunctions.

In this respect, similar results were documented on different animal species treated with Nano-Se (Qin *et al.*, 2016; Halawa *et al.*, 2023b; Liu *et al.*, 2024).

### **Thyroid hormones and antioxidant status of ewes and their suckling lambs:**

Profiles of serum thyroid hormone concentrations and antioxidant status of the ewes and their respective lambs are illustrated in Table (7). The data showed that serum thyroid hormone concentrations ( $T_3$  and  $T_4$ ) were higher ( $P < 0.05$ ) for both ewes fed T1 (Sodium selenite) or T2 (Nano-Se) and their lambs when compared to their respective control. The levels of  $T_3$  and  $T_4$  hormones were also noticed to be increased ( $P < 0.05$ ) for ewes fed T2 and their suckling lambs compared to those fed T1. Meanwhile, the values of the  $T_3/T_4$  ratio did not change among treatments. These findings were consistent with those of Salam *et al.*, (2021) who reported that giving pregnant ewes Nano-Se at 0.1 and 0.2 mg / kg DM increased  $T_3$  and  $T_4$  of their suckling lambs by 36.25 and 45.31; 2.08 and 9.4 %, respectively based on the control. A similar trend of increasing  $T_3$  levels, but not for  $T_4$  or  $T_4/T_3$  ratio, due to vitamin E plus Se supplement was detected in suckling lambs (Ibrahim, 2017). The enhancement in thyroid hormones levels could be explained by the fact that Se serving as a critical structural element in selenocysteine, which exists in the active sites of iodothyronine deiodinase and thioredoxin reductases enzymes, which are involved in thyroid hormones metabolism (Adadi *et al.*, 2019; Arshad *et al.*, 2021).



**Table 6: Effects of inorganic Se or Nano-Se dietary supplementation on some serum biochemical indices of Ossimi ewes and their lambs (Mean  $\pm$  SEM).**

Parameters	Treatments			± SEM
	Control	T1	T2	
Ewes:				
Total protein (g/dl)	6.37 <sup>b</sup>	7.31 <sup>a</sup>	7.61 <sup>a</sup>	0.28
Albumin (g/dl)	4.12 <sup>b</sup>	4.53 <sup>a</sup>	4.68 <sup>a</sup>	0.18
Globulin (g/dl)	2.25 <sup>b</sup>	2.78 <sup>a</sup>	2.93 <sup>a</sup>	0.17
A/ G ratio	1.83	1.63	1.58	0.08
Glucose (mg/dl)	78.54	79.17	79.39	1.58
Cholesterol (mg/dl)	100.20 <sup>a</sup>	93.81 <sup>b</sup>	90.73 <sup>b</sup>	1.84
AST (U/L)	54.67 <sup>a</sup>	46.50 <sup>b</sup>	30.17 <sup>c</sup>	1.93
ALT (U/L)	28.66 <sup>a</sup>	26.50 <sup>b</sup>	22.33 <sup>c</sup>	0.62
Urea (mg/dl)	67.74 <sup>a</sup>	61.25 <sup>b</sup>	45.59 <sup>c</sup>	1.62
Suckling lambs:				
Total protein (g/dl)	6.13 <sup>b</sup>	7.09 <sup>a</sup>	7.61 <sup>a</sup>	0.12
Albumin (g/dl)	3.26 <sup>b</sup>	3.74 <sup>a</sup>	4.68 <sup>a</sup>	0.07
Globulin (g/dl)	2.88 <sup>b</sup>	3.35 <sup>a</sup>	2.93 <sup>a</sup>	0.07
A/ G ratio	1.14	1.12	1.58	0.03
Glucose (mg/dl)	75.53	76.65	79.39	5.06
Cholesterol (mg/dl)	97.72 <sup>a</sup>	90.12 <sup>b</sup>	90.73 <sup>b</sup>	1.83
AST (U/L)	45.67 <sup>a</sup>	42.83 <sup>b</sup>	30.17 <sup>c</sup>	1.59
ALT (U/L)	26.22 <sup>a</sup>	22.06 <sup>b</sup>	22.33 <sup>c</sup>	1.45
Urea (mg/dl)	76.94 <sup>a</sup>	59.17 <sup>b</sup>	45.59 <sup>c</sup>	3.38

<sup>a,b,c</sup> means within the same row having different superscripts significantly different (\* P<0.05),

Cont.: control, T1: Sodium selenite (0.3 mg / kg DM), T2: Nano selenium (0.3 mg /kg DM)

AST = aspartate transaminase

ALT = alanine transaminase

Therefore, the current findings suggest that dietary Nano Se supplementation may have enhanced metabolic functions in Ossimi ewes and their suckling lambs, potentially explaining the observed increases in nutrients digestibility and overall production efficiency.

Additionally, other data in Table (7) showed that serum total antioxidant capacity (TAC) and glutathione-S-transferase (GST) levels were higher (P<0.05) for ewes fed T1 or T2 and their respective lambs when compared to control. It was observed that the values of TAC and GST concentrations increased

(P<0.05) for ewes fed T2 and their lambs compared to those fed T1. This increase in antioxidant levels could be explained by the ability of nano particles to trap free radicals with a greater antioxidant effect and have an increased adsorptive ability due to interactions between the nanoparticles and NH, C=O, COO-, and C-N functional groups of proteins (Zhang *et al.* 2007; Vajpeyee *et al.*, 2024). The significant increase of GST and TAC concentrations may indicate that maternal Se supplementation at late pregnancy and the suckling period could enhance the antioxidant



status of Ossimi ewes and their respective lambs. These findings are in the same line with the previous findings which indicated that Se supplementation increased TAC concentrations

(Sushma *et al.* 2015; Shi *et al.* 2018; Abdel-Raheem *et al.*, 2019; Vajpeyee *et al.*, 2024), and GST in small ruminant (Mohamed *et al.*, 2017; Szeligowska *et al.*, 2022).

**Table 7: Effects of inorganic Se or Nano-Se dietary supplementation on thyroid hormone concentrations and serum antioxidant status of Ossimi -ewes and their lambs (Mean  $\pm$  SEM).**

Parameters	Treatments			± SEM
	Control	`T1	T2	
Ewes:				
T <sub>3</sub> (nmol/l)	1.98 <sup>c</sup>	2.33 <sup>b</sup>	2.75 <sup>a</sup>	0.04
T <sub>4</sub> (nmol/l)	38.95 <sup>c</sup>	45.85 <sup>b</sup>	55.61 <sup>a</sup>	1.27
T <sub>3</sub> /T <sub>4</sub>	0.05	0.05	0.05	0.01
TAC (mM/L)	1.58 <sup>c</sup>	2.14 <sup>b</sup>	2.73 <sup>a</sup>	0.06
GST (U/ml)	90.31 <sup>c</sup>	127.01 <sup>b</sup>	148.47 <sup>a</sup>	3.26
Suckling lambs:				
T <sub>3</sub> (nmol/l)	2.02 <sup>c</sup>	2.49 <sup>b</sup>	3.06 <sup>a</sup>	0.06
T <sub>4</sub> (nmol/l)	46.68 <sup>c</sup>	60.21 <sup>b</sup>	75.42 <sup>a</sup>	1.98
T <sub>3</sub> /T <sub>4</sub>	0.043	0.041	0.041	0.01
TAC (mM/L)	1.95 <sup>c</sup>	2.73 <sup>b</sup>	3.36 <sup>a</sup>	0.07
GST (U/ml)	58.84 <sup>c</sup>	105.62 <sup>b</sup>	144.20 <sup>a</sup>	3.18

<sup>a,b,c</sup>, means within the same row having different superscripts significantly different (\* P<0.05),

Cont.: control, T1: Sodium selenite (0.3 mg / kg DM), T2: Nano selenium (0.3 mg /kg DM)

TAC = total antioxidant capacity

GST = glutathione-S- transferase

## Conclusion

Based on the present study, it could be concluded that dietary supplementation of Nano-Se at 0.3 mg/Kg DM to Ossimi ewes, at late gestation stage and the suckling period, was more potent than inorganic Se (sodium selenite) to improve their nutrients digestibility, productive performance, some serum biochemical indices and antioxidant status, leading to better growth performance and health of their suckling lambs.

## Ethics statement

All animal procedures were approved by the Animal Care and Use Committee of Faculty of Agriculture, Minia University and were in accordance with the university's guidelines for animal research (MU FA 0311123).

## REFERENCES

- Abdel Ghfar, S. S.; M. E. Ali; M. A. Momenah; F. A. Al-Saeed; A. A. Al-Doaiss; Y. S. Mostafa; A. E. Ahmed and M. Abdelrahman (2022): Effect of *Allium sativum* and *Nigella sativa* on alleviating aluminum toxicity state in the albino rats. *Front Vet Sci.*, 9:1042640.
- Abdel-Raheem, S. M.; G. B. Mahmoud; W. Senosy and T. M. El-Sherry (2019): Influence of vitamin E and selenium supplementation on the performance, reproductive indices and metabolic status of Ossimi ewes. *Slovenian Veterinary Research/Slovenski Veterinarski Zbornik*, 56.



- Abdelrazek, S. and R. Fayed (2025): Assessment of Selenium in Agrosystem of the Northwest Delta, Egypt. *Alexandria Science Exchange Journal*, 46(1): 1-15.
- Adadi, P., N. V. Barakova; K. Y. Muravyov and E. F. Krivoschapkina (2019): Designing selenium functional foods and beverages: A review. *Food Res Int.*, 120: 708e25.
- Aghwan, Z. A.; A. Q. Sazili; A. R. Alimon; Y. M. Goh and M. Hilmi (2013): Blood haematology, serum thyroid hormones and glutathione peroxidase status in Kacang goats fed inorganic iodine and selenium supplemented diets. *Asian-Australasian J. Anim. Sci.*, 26: 1577–82.
- Alahmadi, B. A.; S. H. El-Alfy; A. M. Hemaïd and I. M. Abdel-Nabi (2020): The protective effects of vitamin E against selenium-induced oxidative damage and hepatotoxicity in rats. *J. Taibah. Univ. Sci.*, 14: 709–720.
- Alimohamady, R.; H. Aliarabi; A. Bahari. and A. H. Dezfoulan (2013): Influence of different amounts and sources of selenium supplementation on performance, some blood parameters, and nutrient digestibility in lambs. *Biological Trace Element Research*, 154, 45-54.
- Aljaf, K. A.; M. N. Bolshakova (2023): Nano-selenium-mediated alterations in lipid profile, liver and renal functions, and protein parameters in male lambs: An experimental study. *RUDN J Agron Anim Ind.*, 18 (2), 230-240.
- AOAC (2019): Official Methods of Analysis of AOAC international. Association of Analytical Chemists. 21st ed. Rockville, MD., USA.
- Arshad, M.; H. M. Ebeid and F. Hassan (2021): Revisiting the effects of different dietary sources of selenium on the health and performance of dairy animals: a review. *Biol. Trace Elem. Res.*, 199:3319 - 3337
- Ashmawy, B. M. (1980): Comparison of three techniques for the estimation of the milk production of small ruminants. *Egyptian J. Anim. Prod.*, 20: 11- 16.
- Bagnicka, E.; E. M. Kościuczuk; J. Jarczak; A. Jóźwik; N. Strzałkowska; D. Słoniewska and J. Krzyżewski (2017): The effect of inorganic and organic selenium added to diets on milk yield, milk chemical and mineral composition and the blood serum metabolic profile of dairy cows. *Anim Sci Paper Rep.*, 35(1):17–33
- Barcelos, B.; V. Gomes; A. M. Vidal; J. E. de Freitas; M. L. de Araújo; H. D. Alba and A. S. Netto (2022): Effect of selenium and vitamin E supplementation on the metabolic status of dairy goats and respective goat kids in the peripartum period. *Trop. Anim. Health Prod.*, 54, 1–13
- Barcelos, B.; V. Gomes; A. M. Vidal; J. E. de Freitas; M. L. de Araújo; H. D. Alba and A. S. Netto (2023): Milk yield, composition and immune status of dairy goats and respective goat kids fed diets with selenium and vitamin E supplementation. *Small Ruminant Research*, 225: 106999.
- Bolshakova, M.; K. A. H. Aljaf (2022): The effect of dietary nano-selenium and cannabis seeds on liver tissues and functions in male Karadi lambs. *J Pharm Negat Results*, 13 (5), 1051-1057.
- Budak, D. (2024): Effects of Nano Selenium on Some Metabolic and Rumen Parameters in Dorper Sheep. *Van Vet J.*, 35 (1), 83-88.
- Chen, J.; F. Zhang; W. Guan; H. Song; M. Tian, L. Cheng; K. Shi; J. Song; F. Chen; S. Zhang; F. Yang; C. Ren and Y. Zhang (2019): Increasing selenium supply for heat-stressed or actively cooled sows improves piglet



- preweaning survival, colostrum and milk composition, as well as maternal selenium, antioxidant status and immunoglobulin transfer. *J Trace Elem. Med. Biol.*, 52: 89-99.
- Cui, X., Z. Wang; Y. Tan; S. Chang; H. Zheng; H. Wang; T. Yan; T. Guru and F. Hou (2021): Selenium yeast dietary supplement affects rumen bacterial population dynamics and fermentation parameters of Tibetan sheep (*Ovis aries*) in alpine meadow. *Front Microbiol*, 12, 663945.
- Duncan, D. B. (1955): Multiple range test and multiple F-test. *Biometrics*, 11: 1-42.
- Economides, S. and A. Louca (1981): The effects of the quality and quantity of feed on the performance of pregnant and lactating goats. *Proc. Int. Symp. Nutr. Syst. Goat Feeding*, Tours, France, 286-291.
- Ferrari L.; D. Cattaneo; R. Abbate; M. Manoni; M. Ottoboni; A. Luciano; C. Holst and L. Pinotti (2023): Advances in selenium supplementation: From selenium-enriched yeast to potential selenium-enriched insects, and selenium nanoparticles. *Animal Nutrition*, 14:193e203
- Gaafar, H.; M. El-Nahrawy; M. El-Gendy; K. El-Riedy; M. Zommara; R. Mesbah and M. Ghanimah (2021): Nutritional effect of different forms of selenium additive on productive performance of dairy Zaraibi goats and their suckling kids. *Indian J. Vet. Res*, 30(2), 10-19.
- Gangadoo, S.; I. Dinev; N. L. Willson; R. J. Moore; J. Chapman and D. Stanley (2020): Nanoparticles of selenium as high bioavailable and non-toxic supplement alternatives for broiler chickens. *Environmental Science and Pollution Research*, 27, 16159-16166.
- Ghaderzadeh, S. (2016): Effects of dietary protected conjugated linoleic acid (CLA) and different levels of Nano-Se on performance, some parameters of immune system fatty acids profiles of tissues and related genes in tissue and blood of male Moghani lambs.' PhD Thesis, University of Mohaghegh Ardabili, Ardabil, Iran
- Goff, J. P. (2018): Invited review: Mineral absorption mechanisms, mineral interactions that affect acid-base and antioxidant status, and diet considerations to improve mineral status. *Journal of Dairy Science*, 101(4), 2763-2813.
- Goring, H. K. and P. J. Van Soest (1970): Forage fiber analysis U.S.D.A. Agricultural handbook No. 379.
- Gu, X. and C. Gao (2022): New horizons for selenium in animal nutrition and functional foods. *Animal Nutrition*, 11: 80e6.
- Haenlein, G. F. W. and M. Anke (2011): Mineral and element research ingots: a review. *Small Rumin. Res.*, 95(1): 2-19.
- Halawa, E.; T. Imbabi; O. Farid; A. Radwan and A. ElSayed (2023a): The influence of selenium nanoparticles and L-Carnitine on various biochemical markers and oxidative stress status in Ossimi ewes during post-partum periods. *Benha Veterinary Medical Journal*, 44 (1), 34-38.
- Halawa, E.; T. Imbabi; O. Farid; A. Radwan and A. ElSayed (2023b): Ameliorating Effect of Selenium Nanoparticles and L-Carnitine on Some Haemato-Biochemical Parameters and Oxidative Stress Status During Pregnancy Periods in Ossimi Ewes. *Annals of Agricultural Science Moshtohor*, 61(1): 49 – 58.
- Hall, J. A.; W. R. Vorachek; W. C. Stewart; M. E. Gorman; W. D. Mosher; G. J. Pirelli and G. Bobe (2013): Selenium supplementation restores innate and humoral immune responses in foot rot affected sheep. *PLOS One*, 8: p.e82572.



- Han, L. Q.; K. Pang; T. Fu; C. J. C. Phillips and T. Y. Gao (2021): Nano- selenium supplementation increases selenoprotein (Sel) gene expression profiles and milk selenium concentration in lactating dairy cows. *Biol. Trace Elem. Res.*, 199 (1):113–119.
- Hashem, N. M. and S Z. EL –Zarkouny (2016): Postpartum associated metabolism, milk production and reproductive efficiency of Barki and Rahmani subtropical fat -tailed breeds. *Asian J. Anim. Vet. Adv.* 11, 184–189.
- Hendawy, A. O.; S. Sugimura; K. Sato; M M. Mansour; A. H. Abd El-Aziz; H. Samir; M. A. Islam; A. B. M. R. Bostami; A S. Mandour; A. Elfadadny, A.; R. Ragab; H. Abdelmageed and A. Ali (2022): Effects of selenium supplementation on rumen microbiota, rumen fermentation, and apparent nutrient digestibility of ruminant animals: a review. *Fermentation*, 8: 4.
- Hofstee, P.; D. R. McKeating; L. A. Bartho; S. T. Anderson; A. V. Perkins and J. S. M. Cuffe (2020): Maternal selenium deficiency in mice alters offspring glucose metabolism and thyroid status in a sexually dimorphic manner. *Nutrients*, 12: 267.
- Ibrahim, E. M. (2017). Effect of parenteral supplementation of vitamin E plus selenium on nutrient digestibility, productive performance and some serum biochemical indicators of lambs. *Egyptian Journal of Sheep and Goats Sciences*, 12(1): 1-12.
- Ibrahim, E. M. and M. Y. Mohamed (2018): Effect of different dietary selenium sources supplementation on nutrient digestibility, productive performance and some serum biochemical indices in sheep. *Egyptian Journal Nutrition and Feeds*, 21: 53–64.
- Kaneko, J.J.; J. W. Harvey and M. L. Bruss (2008): *Clinical Biochemistry of Domestic Animals*, 6th ed.; Elsevier Academic Press: Amsterdam, The Netherlands, p. 931.
- Kareem, O. M.; A. M. Saadik and O. D. Almallah (2024): Addition of selenium nanoscale to high diets with wheat bran and its effect on milk production and composition in Awassi ewes. *Assiut Vet. Med. J. Vol.* (70): 323-334.
- Kasim, H.W.; M. N. Abdullah; I. H. Hamad and O. D. Almallah (2023): Effect of Adding Selenium Vitamin E to Feed Containing Acetic Acid on Milk Production and its Fat Content in Awassi Ewes. In *IOP Conference Series: Earth and Environmental Science* (Vol. 1213, No. 1, p. 012073). IOP Publishing.
- Kieliszek, M. and S. Blazejak (2016): Current knowledge on the importance of selenium in food for living organisms: a review. *Molecules* 21, 609.
- Kumar, S.; S. Vaswani; V. Kumar; M. Anand; M., Kumar; R. Kushwaha; A. Kumar and S. P. Singh (2022): Effect of dietary supplementation of different sources of selenium on growth performance and nutrient utilization of Barbari bucks. *J. Anim. Res.*, 12 (06): 949-955.
- Lee, M. R. F.; H. R. Fleming; T. Cogan; C. Hodgson and D. R. Davies (2019): Assessing the ability of silage lactic acid bacteria to incorporate and transform inorganic selenium within laboratory scale silos. *Anim. Feed Sci. Technol.*, (253): 125–134.
- Lepherd, M. L.; P. J. Canfield; G. B. Hunt and K. L. Bosward (2009): Haematological, biochemical and selected acute phase protein reference intervals for weaned female Merino lambs. *Aust. Vet. J.*, 87: 5–11.
- Li, L.P.; L. Qu and T. Li (2022): Supplemental dietary Selenohomolanthionine affects growth and rumen bacterial population of



- Shaanbei white cashmere wether goats. *Front Microbiol*, 13: 942848.
- Liu, Y.; C. Wang; Q. Liu; G. Guo; W. Huo; Y. Zhang; C. Pei; S. Zhang and J. Zhang (2019): Effects of sodium selenite addition on ruminal fermentation, microflora and urinary excretion of purine derivatives in Holstein dairy bulls. *J. Anim. Physiol. Anim. Nutr. (Berl.)* 103: 1719–1726.
- Liu, Y.; J. Zhang; L. Bu; W. Huo; C. Pei and Q. Liu (2024): Effects of nanoselenium supplementation on lactation performance, nutrient digestion and mammary gland development in dairy cows. *Animal Biotechnology*, 35 (1): 2290526.
- Malyugina, S.; S. Skalickova; J. Skladanka; P. Slama and P. Horky (2021): Biogenic selenium nanoparticles in animal nutrition: a review. *Agriculture*, 11: 1244.
- Milewski, S.; P. Sobiech; J. Błaz' ejak-Grabowska; R. Wójcik; K. Zarczyn' ska; J. Micin' ski and K. Za' bek (2021): The efficacy of a long-acting injectable selenium preparation administered to pregnant ewes and lambs. *Animals*, 11: 1076.
- Mohamed, M. Y.; E. M. M. Ibrahim and A. M. Abd El-Mola (2017): Effect of selenium yeast and/or vitamin E supplemented rations on some physiological responses of post-lambing Ossimi ewes under two different housing systems. *Egyptian Journal of Nutrition and Feeds*, 20 (3): 361-378.
- Mousaie, A.; R. Valizadeh; A. A. Naserian; M. Heidarpour and H. Mehrjerdi (2014): Impacts of feeding selenium-methionine and chromium-methionine on performance, serum components, antioxidant status and physiological responses to transportation stress of Baluchi ewe lambs. *Biological Trace Element Research*, 162, 113- 123.
- Mudgal, V.; A. K. Garg; R. S. Dass and V. P. Varshney (2008): Effect of selenium and copper supplementation on blood metabolic profile in male buffalo (*Bubalus bubalis*) calves. *Biol Trace Elem Res.*, 121(1):31–38.
- NRC (2007): 'Nutrient requirements of small ruminants, sheep, goats, cervids, and new world camelids. (National Academies Press: Washington, DC).
- Pan, Y.; Y. Wang; S. Lou; M. Wanapat; Z. Wang; W. Zhu and F. Hou (2021): Selenium supplementation improves nutrient intake and digestibility, and mitigates CH<sub>4</sub> emissions from sheep grazed on the mixed pasture of alfalfa and tall fescue. *J. Anim. Physiol. Anim. Nutr.*, 105: 611–620.
- Pecoraro, B. M.; D. F. Leal; A. Frias-De-Diego; M. Browning; J. Odle and E. Crisci (2022): The health benefits of selenium in food animals: a review. *J Anim Sci Biotechnol.*, 13(1):58.
- Qin, F.; F. Chen; F. H. Zhao; T. M. Jin and J. Ma (2016): Effects of nanoselenium on blood biochemistry, liver antioxidant activity and GPx-1 mRNA expression in rabbits. In: *International Conference on Biomedical and Biological Engineering*. Atlantis Press. Pp. 166-171.
- Rabee, A. E.; M. M. Khalil; G. A. Khadiga; A. Elmahdy; E. A. Sabra; M. A. Zommara, and I. M. Khattab (2023): Response of rumen fermentation and microbiota to dietary supplementation of sodium selenite and bio-nanostructured selenium in lactating Barki sheep. *BMC Veterinary Research*, 19(1), 247.
- Rajab, W.; M. Rudiansyah; M. M. Kadhim; A. Shamsiev; S. Prakaash; M. Lafta; S. Aravindhana and Y. Mustafa (2023): The Role of Selenium on the Status of Mineral Elements and Some Blood Parameters of Blood Serum of Lambs. *Archives of Razi Institute*, 78(1): 135–144.
- Salam, A. Y.; I. S. EL-Shamaa; A. M. Metwally; A. Y. El-Hewaty; T. A. Mahmoud and M. A.



- Zommara (2021): Effect of selenium administration on reproductive outcome and biochemical parameters to ewes and their lambs. *J. of Animal and Poultry Production*, Mansoura Univ., Vol. 12 (12):379 – 386.
- Saleh, A. A. and T. A. Ebeid (2019): Feeding sodium selenite and nano-selenium stimulates growth and oxidation resistance in broilers. *South African Journal of Animal Science* 49: 176–84.
- Salles, M. S.; T. S. Sam´ora; A. M. Della Libera; A. S. Netto; L. C. R. Junior; M. G. Blagitz and J. E. de Freitas (2022): Selenium and vitamin E supplementation ameliorates the oxidative stress of lactating cows. *Livestock Sci.*, 255: 104807.
- SAS (2012): SAS/STAT User’s Guide: Statistics. SAS Institute Inc., Cary. NC., USA.
- Shams, A. S.; M. A Zommara; M. E. Sayed-Ahmed and M. M. El-Nahrawy (2020): Growth performance and immunity response of suckling Friesian calves fed on ration supplemented with organic or nano selenium produced by lactic acid bacteria. *Egyptian Journal of Nutrition and Feeds*, 23(2), 205-217.
- Shi, L.; W. Xun; W. Yue; C. Zhang; Y. Ren; Q. Liu; Q. Wang and L. Shi (2011): Effect of elemental nano-selenium on feed digestibility, rumen fermentation, and purine derivatives in sheep. *Animal Feed Science Technology* 163: 136–42.
- Shi, L.; Y. Ren; C. Zhang; W. Yue and F. Lei (2018): Effects of organic selenium (Se-enriched yeast) supplementation in gestation diet on antioxidant status, hormone profile and haemato-biochemical parameters in Taihang Black Goats. *Animal Feed Science Technology* 238: 57–65.
- Silveira, R.; B. Silva; A. Vasconcelos; D. Façanha; T. Martins; M. Rogério and J. Ferreira (2021): Does organic selenium supplement affect the thermoregulatory responses of dairy goats? *Biol. Rhythm. Res.*, 52: 1–13.
- Sushma, K; Y R. Reddy; N. N. Kumari; P. B. Reddy; T. Raghunandan and K. Sridhar (2015): Effect of selenium supplementation on performance, cost economics, and biochemical profile of Nellore ram lambs. *Veterinary World*, 8: 1150–55.
- Szeligowska, N.; P. Cholewińska; J. Smoliński; K. Wojnarowski; P. Pokorny; K. Czyż and K. Pogoda-Sewerniak (2022): Glutathione S-transferase (GST) and cortisol levels vs. microbiology of the digestive system of sheep during lambing. *BMC Veterinary Research*, 18(1): 107-114.
- Tian, X.; P. Paengkoum; S. Paengkoum; S. Thongpea and C. Ban (2018): Comparison of forage yield, silage fermentative quality, anthocyanin stability, antioxidant activity, and in vitro rumen fermentation of anthocyanin-rich purple corn (*Zea mays* L.) stover and sticky corn stover. *J. Integr. Agr.*,17: 2082–2095.
- Tian, X.; X. Wang; J. Li; Q. Luo; C. Ban and Q. Lu (2022): The Effects of selenium on rumen fermentation parameters and microbial metagenome in Goats. *Fermentation*, 8: 240.
- Ucar, S. K.; M. Coker; E. Sozmen; D. G. Simsek and S. Darcan (2010): An association among iron, copper, zinc, and selenium, and antioxidative status in dyslipidemic pediatric patients with glycogen storage disease types of IA and III. *J Trace Elem Med Biol.*, 24(1): 42–45.
- Ullah, H.; R. U. Khan; M. Mobashar; S. Ahmed; A. Sajid; N. Khan; T. Usman; I. Khattak, and H. Khan (2019): Effect of yeast-based selenium on blood progesterone, metabolites and milk yield in Achai dairy cows. *Ital J Anim Sci.* 18(1): 1445–1450.



- Vajpeyee S.; J. Ramesh; R. Karunakaran; J. Muralidharan; V. Sankar and K. Das (2024): Effect of different selenium sources on nutrient digestibility, performance and antioxidant status in Mecheri lambs. *The Indian Journal of Animal Sciences*, 94(2), 161–165.
- Van Keulen, J. and B. A. Young (1977): Evaluation of acid insoluble ash as a natural marker in ruminant digestibility studies. *J. Anim. Sci.*, 44: 282.
- Vasil, M.; F. Zigo; J. Elečko; M. Zigová and Z. Farkašová (2017): Effect of peroral supplementation with selenium and vitamin E during late pregnancy on udder health and milk quality in dairy cows. *Potravinárstvo Slovak J Food Sci.*, 11(1):535–538
- Wang, Z. F.; Y. H. Tan; X X. Cui; S. H. Chang; X. Xiao; T. H. Wang and F. J. Hou (2019): Effect of different levels of selenium yeast on the antioxidant status, nutrient digestibility, selenium balances and nitrogen metabolism of Tibetan sheep in the Qinghai-Tibetan Plateau. *Small Rumin. Res.*, 180: 63–69.
- Yaghmaie, P.; A. Ramin; S. Asri-Rezaei and A. Zamani (2017): Evaluation of glutathione peroxidase activity, trace minerals and weight gain following administration of selenium compounds in lambs. *Veterinary Research Forum* 8: 133–37.
- Zhang, C X.; J. B. Yue; X. F. Zhang; W. F. Dong and F. L. Lei (2007): Effect of sodium selenite on growth performance, blood physiology index and biochemistry characteristics in weaned kids. *Chinese Journal of Animal Science* 43: 36–39.
- Zhang, Z. D.; C. Wang; H. S. Du; Q. Liu; G. Guo; W. J. Huo; J. Zhang; Y. L. Zhang; C. X. Pei and S. L. Zhang (2020): Effects of sodium selenite and coated sodium selenite on lactation performance, total tract nutrient digestion and rumen fermentation in Holstein dairy cows. *Animal.*, 14(10):2091–2099.



### الملخص العربي

الأداء الإنتاجي وبعض المؤشرات البيوكيميائية للنعاج الأوسيمي وحملاتها للإمداد الغذائي بالسلينيوم

عمادالدين محمد إبراهيم<sup>١</sup>، محمود يس محمد<sup>٢</sup>

<sup>١</sup>قسم الإنتاج الحيواني، كلية الزراعة، جامعة المنيا، المنيا، مصر

<sup>٢</sup>معهد بحوث الإنتاج الحيواني، مركز البحوث الزراعية، وزارة الزراعة، مصر

استخدم في هذه الدراسة عدد أربعة وعشرون من النعاج الأوسيمي العشار لتقييم تأثير الإمداد الغذائي بالسلينيوم الغير عضوي والنانو سلينيوم على الأداء الإنتاجي، معاملات الهضم الغذائية، بعض مؤشرات السيرم البيوكيميائية و حالة مضادات الأكسدة للنعاج الأوسيمي وحملاتها الرضيعة. غذيت النعاج على العليقة الأساسية (الكنترول) مضافاً إليها ٠.٣ ملجم سلينيوم / كجم مادة جافة في صورة سليينات صوديوم (T1) أو نانو سلينيوم (T2) .

أظهرت النتائج أن قيم معاملات هضم كلا من المادة العضوية، البروتين الخام، المستخلص الأثيري، الألياف الخام و الألياف المستخلصه بالمحاليل الحامضية ارتفعت معنوياً ( $P<0.05$ ) للنعاج المغذاه على المعامله (T2) مقارنة بتلك المغذاه على الكنترول أو المعاملة (T1). زادت متوسطات وزن الميلاد، وزن الجسم النهائي ومعدل النمو اليومي معنوياً ( $P<0.05$ ) للحملان المولودة للنعاج المغذاه على المعاملات (T1) أو (T2) مقارنة بالكنترول على الترتيب. سجلت النعاج التي غذيت على المعاملات (T1) أو (T2) قيماً أعلى معنوياً ( $P<0.05$ ) في إنتاج اللبن، محصول اللبن المعدل لنسبة الدهن، محصول دهن اللبن ومحصول بروتين اللبن مقارنة بتلك المغذاه على الكنترول. كان تركيز السيرم من الأمينوجلوبيولين، البروتينات الكلية، الألبومين، الجلوبيولين، هرمونات الغدة الدرقية، قدره الكلية المضاده للأكسدة وكذلك إنزيم الجلوتاثيون – إس- ترانسفيريز أعلى معنوياً ( $P<0.05$ ) لكلا من النعاج المغذاه على المعاملات (T1) أو (T2) وحملاتها الرضيعة مقارنة بتلك المغذاه على الكنترول. وقد حدث انخفاض معنوياً ( $P<0.05$ ) في تركيزات السيرم من الكولستيرول، نشاط انزيمات الكبد (ALT، AST) وتركيز اليوريا لكلا من النعاج المغذاه على المعاملات (T1) أو (T2) وتوابعها من الحملان مقارنة بتلك المغذاه على العليقة الكنترول.

نستخلص من هذه الدراسة أن الإمداد الغذائي بالنانو سلينيوم بتركيز ٠.٣ ملجم / كجم مادة جافة، خلال الفترة الأخيرة من الحمل وفترة الرضاعة، كان أكثر فاعلية من السلينيوم غير العضوي (سليينات الصوديوم) في تحسين معاملات الهضم الغذائية، الأداء الإنتاجي وبعض مؤشرات السيرم البيوكيميائية وكذلك حالة مضادات الأكسدة للنعاج الأوسيمي مؤدياً إلى أداء نمو وحالة صحية أفضل لحملاتها الرضيعة.